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APPENDICES

APPENDIX A: Preparation of solutions

1. LB broth

Tryptone	10.0	g
NaCl	5.0	g
Yeast extracts	5.0	g

pH is adjusted to 7.0 by 1 M NaOH before making the volume up to 100 ml with dd-H₂O. The broth is finally autoclaved.

2. LB plate with ampicillin/IPTG/X-GAL

LB plate gradient is combined as described in 1. The mixture is supplemented with 0.5 mM IPTG and 80 µg/ml X-GAL before pouring plates. Alternatively, 100 µl of 100 mM IPTG and 20 µl of 50 mg/ml X-GAL can be spread on the surface of an LB-ampicillin plate and allowed to absorb for 30 min at 37°C before use.

3. SOC medium

Tryptone	2.0	g
1 M NaCl	1.0	ml
Yeast extracts	0.5	g
1 M KCl	0.25	ml
2 M Mg ²⁺ stock	1.0	ml
2 M glucose	1.0	ml

Add Tryptone, Yeast extract, NaCl and KCl to 97 ml of distilled water. Stir to dissolve. Autoclave and cool to room temperature. Add 2 M of Mg²⁺ stock and 2 M of glucose, each to a final concentration of 20 mM. Bring to 100 ml with dd-H₂O. Filter the complete medium through a 0.2 µm filter unit. The final pH should be 7.0.

4. Marine broth pH 7.3

Peptone	5.0	g
Yeast extract	1.0	g
Ferric Citrate	0.1	g
NaCl	20.0	g

Adjusted pH to be 7.3 by 1 M HCl and adjusted volume to 1,000 ml by dd-H₂O and autoclave.

5. Coomassie stain solution

Coomassie brilliant blue R-250	0.25	g
Methanol	45.0	ml
Acetic acid	10.0	ml
d-H ₂ O adjust volume to	45.0	ml

6. De-staining solution

Methanol	400	ml
Acetic acid	100	ml
d-H ₂ O adjust volume to	500	ml

7. 1xPBS (Phosphate buffer saline)

Na ₂ HPO	40.72	g
KH ₂ PO ₄	0.12	g
KCl	0.10	g
NaCl	0.40	g

Adjust volume to be 1,000 ml by dd-H₂O and autoclave.

8. Colour reagent

Thiourea	0.75	g
Gracial acetic acid	470	ml
O-toluidine	30	ml

Mix and cover with aluminum foil and store at room temperature.

9. 1xTBE buffer (Tris borate EDTA)

Tris	108	g
Boric acid	55	g
EDTA	9.3	g

Adjust volume to 1,000 ml by d-H₂O and autoclave.

10. Gel loading dye

bromophenol blue	0.25 %	
xylene cyanol FF	0.25 %	
ficoll	15.0 %	

11. 1.5 M Tris-HCl, pH 8.8

Tris (hydroxymethyl)-aminometane 18.17 g

Adjust pH to 8.8 by 1 M HCl and adjust volume to 100 ml with dd-H₂O

12. 0.5 M Tris-HCl, pH 6.8

Tris (hydroxymethyl)-aminometane 6.06 g

Adjust pH to 6.8 by 1 M HCl and adjust volume to 100 ml with dd-H₂O

13. Staining solution

Silver nitrate	1.5	g
Formaldehyde	2.25	ml

Adjust volume to 1,500 ml with dd-H₂O

14. Low Molecular Weight markers (LMW, Amersham Biosciences): consists of

Phosphorylase b, rabbit muscle	97	kDa
Albumin, bovine serum	66	kDa
Ovalbumin, chicken egg white	45	kDa
Carbonic anhydrase, bovine erythrocyte	30	kDa
Trypsin inhibitor, soybean	20	kDa
α-lactalbumin, bovine milk	14.4	kDa

15. 30% Acrylamide and 0.8% bis-acrylamide

Acrylamide	29.2	g
N, N'-methylene-bis-acrylamide	0.8	g

Adjust volume to 100 ml with dd-H₂O

16. 5x Loading buffer

1 M Tris-HCl, pH 6.8	0.6	ml
Glycerol	5.0	ml
10% (w/v) SDS	2.0	ml
2-mercaptoethanol	0.5	ml
1% Bromophenol blue	0.5	g

One part of sample buffer is added to four parts of sample. The mixture is heated for 5 min in boiling H₂O before loading to the gel.

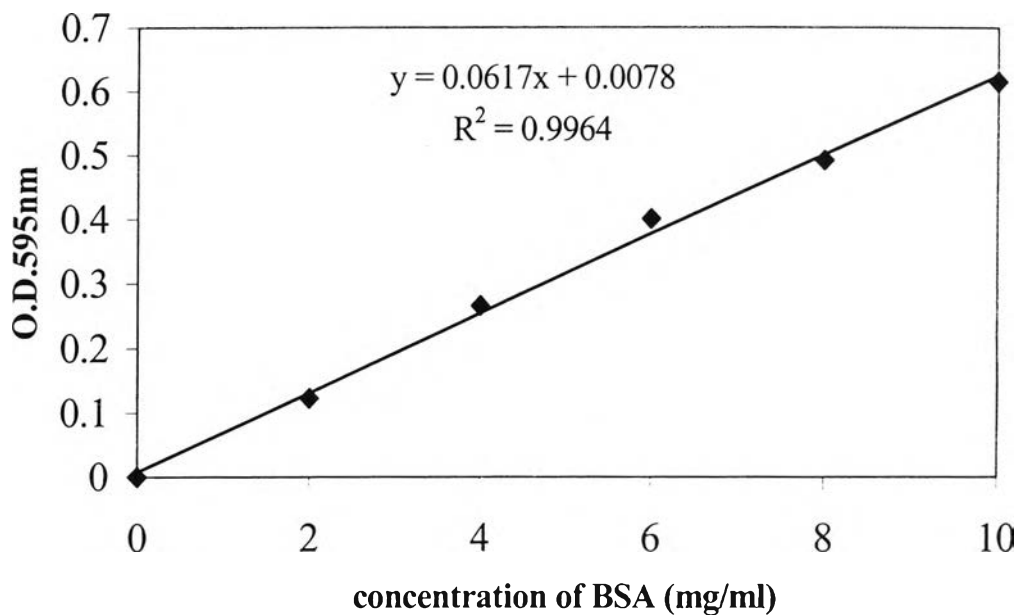
17. Electrophoresis buffer (25 mM Tris, 192 mM glycine and 0.1% SDS)

Tris (hydroxymethyl)-aminometane	6.06	g
Glycine	14.4	g
SDS	1.2	g

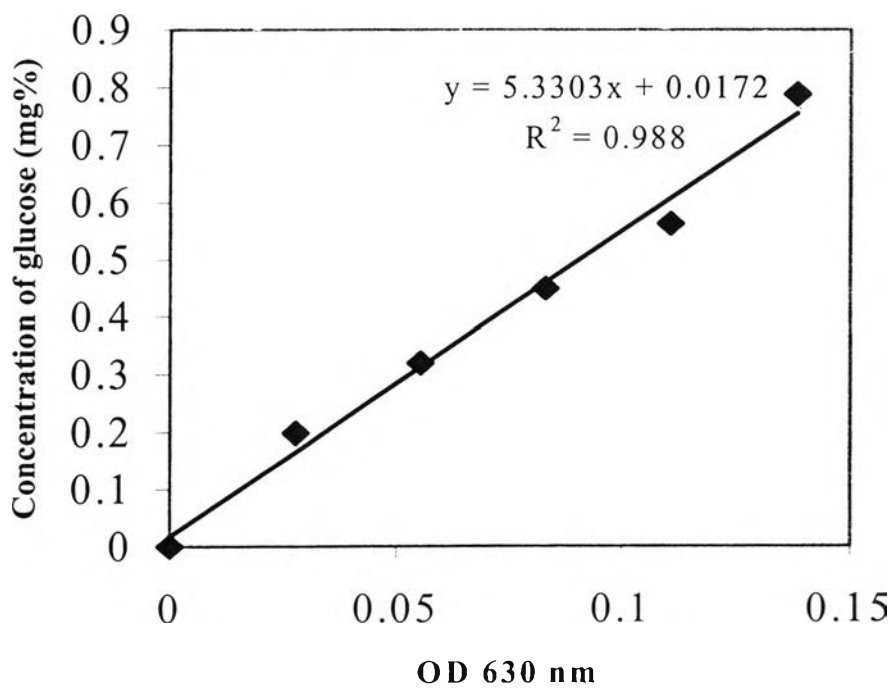
Adjust volume to be 1,000 ml with dd-H₂O

APPENDIX B

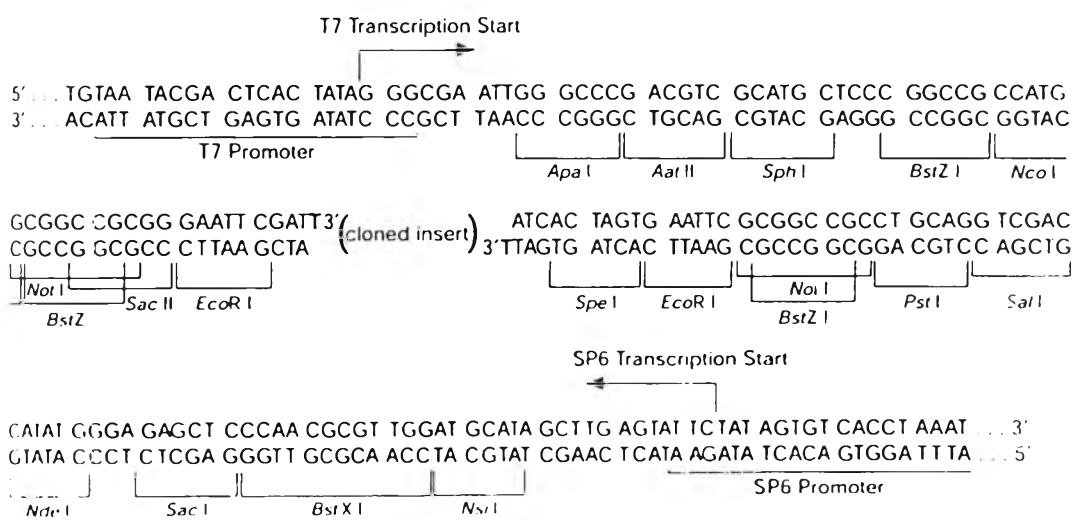
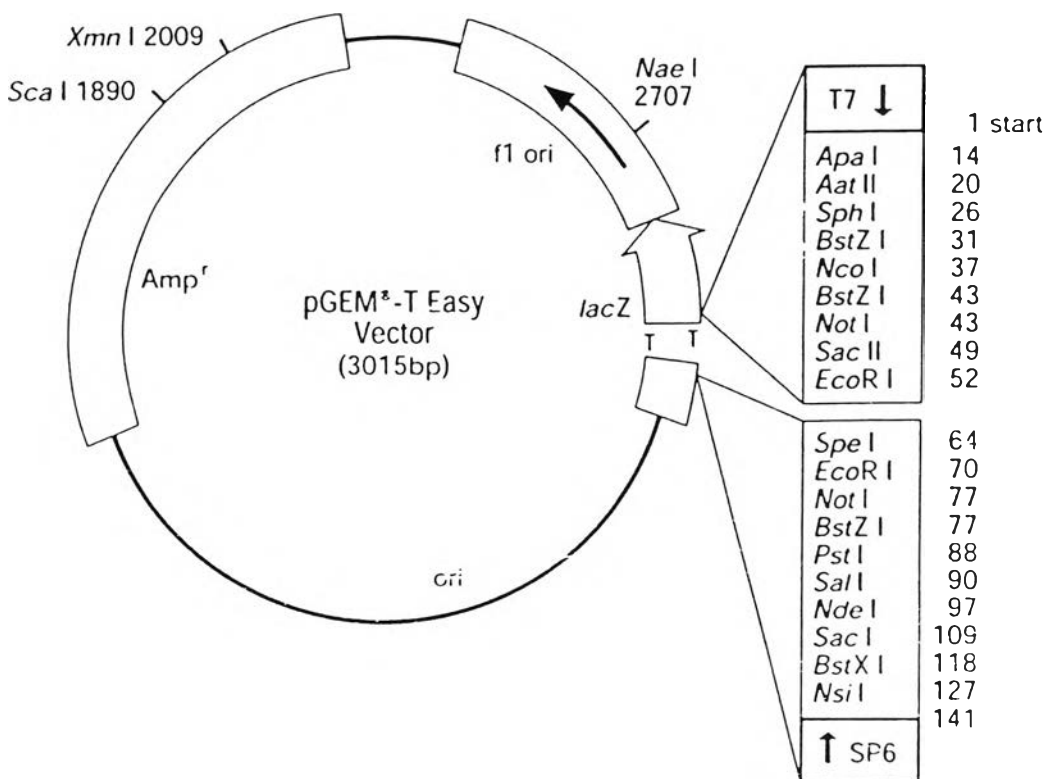
1. Standard curve for protein determination by Bradford's method



2. Standard curve for glucose determination



APPENDIX C

Physical map of pGEM[®]-T easy vector and multiple cloning sites

APPENDIX D

Nucleotides and deduced amino acid sequences

ATTGGGCGATCCGACCACTCTTCACATGTGCAAGGTCCCCTGTTTAGACCATTTCGGACCAGG
 TGCCTGTTTGCAATTTCTGTTTCAAGGACACTATTACCCTGTCCCTTGGGTTTCAGTCTAGG
 ACGGGATCCTGATATTGATTCTGATTCAGGATCCCAACCCTCAGACAGGCCGTGCCAGGTTA
 TCTTTGGCAGCACCCGAACCCTCACGAGGCTTGGTGATATATACATGTGTCCATCCTGACAC
 ATGTTCTTCAGAGGTCAATAGAGCCAAAGATCCTCATCCTTGATTATCACTGAAGGGAAGTC
 GTCGATGACATCCTCTTCACATATGTCTTCATCATAAGCCCCGTTAAGGAACGCTGCAGCCA
 TACAGGTCTGCTCATGT**CCTCGCAGCTT**

Figure D.1 Nucleotide and deduced amino acid sequences of clone 1. Bold letters indicate the regions of primers.

ATTGGGCGAGATCAGTTTTCCGTGTTTTTTTTTCAGAAAATAGGATCAGAAGTCATACATCCGG
 TGTTCTGTCTGACAAGTTACGAGTTTATTGTTCCCTGAATGTAGAGTTAAATTTGTGAATGNA
 CGCCATGCTGAGGAGTGTTGTTTCAGCCACGTAGATTCACGTGTGTGTGTGCGTGTGTGTGTG
 TGTGTGTGTGTGTGTGTGTGTTTGTGTGTGTGTGTGTGTGTGTGTGTGTGTGTGTGTGTGTA
 TGTGTGTATGTGTGTGTGTGTGTGTGTGTGTACATACGTTTGTGTATGTGTAGATATGTGAA
 GGAATGAAACCTACAATGCATGACATGCATGGAGAGAGGAACGAGTTTTGAAAACCTCCTAT
 AGCTTCATCCCCGCCACTCCATTGCCCT**CGCAGCTT**

Figure D.2 Nucleotide and deduced amino acid sequences of clone 2. Bold letters indicate the regions of primers.

AGGCCGCTTAGAAAAGTGAAACAACAAAGCAGACAGTCAACATTCTCAGCCGTTTCGCGCTTC
 AAGACATGTAGATTTTCAGTAGCTCCTTGTAATTGAGGTACGATTCACTTCCTTGCATGTTCGT
 TTACTCCTCGTTTAACTGTTAATACGTTTTCGTGTAATACTAAATAGCTAGTTCTTACTTCCAT
 GTTTTGCTTTCTACATTGTTTTTATGTGAATACTAGTTACTATCGTTTTGCTCTGTCTCAGTG
 AATAACTTGTAATACTTTGATTTGTTTTTCTAACTGTTTTTGCTCTGAAGTAAATACT
 AGTTACTCCTTCGAATAGGACCTTAAACAAACACATTTTTACTCTGCTGTCTCCAGGATTA
 TATCCGTTAAAAGCCCCATTGAGAATGCCGGGAAAAAATGCAAATTTTTTTAACGGTTATT
 TCCACCAGCTAATTTCCCCTTGCCTCT**TAAGCGGCCT**

Figure D.3 Nucleotide and deduced amino acid sequences of clone 3. Bold letters indicate the regions of primers.

ATTGGGCGATCATAGTTTTCCATCCCGTTCTGTATGTTTAGCAGTTCTAAGTCACTTCTGCC
 CATATTGAGATCTTTGTTCAGGGGTATACAAAGGCAGAAACCTTTGTCTACCCTTGCTTCTC
 TTTCCCTCTATCTTTCTATTAACACTAAGTTTTCCAATCCTTTACATTTCATTTTTTTTTAA
 CGGTAGTTTCATGTCTGAGCCACCGTAGTCACAGTATGATACTTAATTGTCGGTTTCATGTT
 GTGATGCTCTTGGAGTGAGTACGTGGTAGGGTCCCCAGTTCCTTTCCACGGAGAGGCATATA
AGCGGCCT

Figure D.4 Nucleotide and deduced amino acid sequences of clone 4. Bold letters indicate the regions of primers.

AGGCCGCTTAGAAAAGTGAAACAACAAAGCAGACAGTCAACATTCTCAGCCGTTTCGCGCTTC
 AAGACATGTAGATTTTCAGTAGCTCCTTGTAATTGAGGTACGATTCACCTCCTTGCATGTCGT
 TTA CT CCTCGTTTAACTGTTAATACGTTTCGTGTA ACTAAATAGCTAGTTCC TACTTCCAT
 GTTTTGCTTTCTACATTGTTTTTATGTGAATAACTAGTTACTATCGTTTTGCTCTGTCAGTG
 AATAACTTGTA ACTACTTTGATTTGTTTTTCTAAACTGTTTTTGCTCTGAAGTAAATAACT
 AGTTACTCCTTCGAATAGGACCTTAAACAAACACATTTTTACTCTGCTGTCCTCCAGGATTA
 TATCCGTTAAAAGCCCCATT CAGAATGCCGGGAAAAAATGCAAATTTTTTTAACGGTTATT
 TCCACCAGCTAATTTCCCTTGCCTCT**TAAGCGGCCT**

Figure D.5 Nucleotide and deduced amino acid sequences of clone 5. Bold letters indicate the regions of primers.

ATTGGGCGATCCGACCACTCTTCACATGTGCAAGGTCCCCTGTTTAGACCATTTCGGACCAGG
 TGCCTGTTTGCAATTTCTGTTTCAAGGACACTATTACCCTGCCCTTTGGGTT CAGTCTAGG
 AAGGAATCCTGATATTGATTCTGATTCAGGATCCCAACCCTCAGACAGGCCATGCCAGGTTA
 TCTTCGGCACGACCCGAACCTTCACGAGGCTTGGTGATATATACATGTGTCCATCCTGACAC
 ATGTTCTTGAGAGGTCAATAGAGCCAAAGATCCTCATCCTTGGTTATCACTGGAGGGAAGTC
 ATCGATGACATCCTCTTCACATATGTCTTCATCATAAGCCCCGGTTTAAGGGAACCGGCCTT
 GGAAGGCCATTACCAGGGTCCGTT CATTGTCT**CGCAGNTT**

Figure D.6 Nucleotide and deduced amino acid sequences of clone 6. Bold letters indicate the regions of primers.

ATTGGGCGATCAAGCCTCAGCAATATCATTAGCATTTGCCTCATATTATCTCTAGTGCCTGA
 AGTTTCTCCTTCATACCTATCTTGGCATTAAAATCTCTCATAATTATTTAATTATTGTATTT
 TCCTCTATCGCAGGCTAAATGTATATCTCCATAGGAGCCCTCTATTTCTTTATCAACCTTGG
 CTACAGGTTGGGACGTAAACTTTAAGTTGTACCTATTGTTTAGTTTTATTGTTACTGTAGCT
 ACTCTCTCGCTTACACAACAGAATTCCACGATATTCTTTTCTAGGCGTTTGTGAACATGTCC
 ATCTCTTGGTACTTTCTGTTCTTCTCCAAGTCTTCAAACCTTCACAGAGGCCCTACAATATCGT
 ATTTAATGAAGGGAAGTTCTCCAAGCAATGCCAGTGAGTTGGATTTCAGTTCT**CATCGCCCAA**
T

Figure D.7 Nucleotide and deduced amino acid sequences of clone 7. Bold letters indicate the regions of primers.

ATTGGGCGATACAAACAGACAAATAGAGTTCCATGAAATACAGAATAAGAAAATAAGAACAT
 TGAAATGTTTGTCCCTTCACTGTCTTTCACAGACTTAAAGAATCTTTTCCTTTGTGCAGCAAC
 TATATGTAGTCATTATATCGAGGTGTTTCATGTCTACAGGTAAATATAGAAAATATAAAAACG
 AAAATGAAAGAAGTGAAAACACCTAACACGTTACCATAGCAATCCAAACGGCTAATGGCAAC
 TCTTCGGGTTGACAATATGGCAGGTGGGCACGGGCATTCTCGATTCACTCACGAATAGA
 AACAGTCTAAGAAGGGGACTATAGCTTCTTAGATAAAAATGGCTGCTGACTGGGGATCTGCA
 GCCGCTACGCTGT**CATCGCCCAAT**

Figure D.8 Nucleotide and deduced amino acid sequences of clone 8. Bold letters indicate the regions of primers.

ATTGGGCGATCAAGCCTCAGCAATATCATTAGCATTTGCCTCATATTATCTCTAGTGCCTGA
 AGTTTCTCCTTCATACCTATCTTGGCATTAAAATCTCTCATAATTATTTAATTATCGTATTT
 TCCTCTATCGCTGGCTAAATGTATATCTCCATAGAAGCCCTCTATTTCTTTATTACTTTGGC
 TACAGGTTGGGACGTAAACTTTAAGTTGTACCTATTGTTTAGTTTTATTGTTACTGTAGCTA
 CTCCCTCGCTTACACAACAGAATTCCACGATATTCTTTTCTAGGCGTTTGTGAACATGTCCA
 TCTTTTAGTACTTTCTGTTCTTCTCCAAGTCTTCAAACCTTCACAGAGGCCCTACAATATCGTA
 TTTAATGAAGGGAAGTTCTCCAAGCAATGCCAGTAAGTTGGATTTCAGTTCT**CATCGCCCAAT**

Figure D.9 Nucleotide and deduced amino acid sequences of clone 9. Bold letters indicate the regions of primers.

ATTGGGCGATGTTCATTTACACGTGAATATGTAAGTGTATTTGGAATTATTCATCCCAGGGTA
 AACTTATTAATTTACAATGATGTAGTTTTTCGACGCGTACTCATTACCGAGTATACCATG
 TTATCAAAATCTACATTTGGCTAATACCTGTATTGGAGCCTTTAATTCCCTGGCCCCNAAA
 AANTGGCTTTCCCGNATGGGTTATATAACCCGCGTTGGGGCTCGTGTANTCATCNGTGCGCG
 CGNGAGAGATGCATCTACGTTACTATTANTCCTTACTATTATCATCATGTACGCGCCTGTAT
 GATCGAGTGCTCGGCCCTCGCGNTCAGTAAAGTAT**CAGCCATACTA**

Figure D.10 Nucleotide and deduced amino acid sequences of clone 10. Bold letters indicate the regions of primers.

ATTGGGCGATCATGATTACAGTCTTACATATATCTGAATTTCCGTAAGATCCTTTAAAAAAA
 ATCTACGGATTTGGTTCGCGAAAATTGAAGTATCTCTATACGGGTATCAGGCAACCCACTTT
 TGCACCTGTATAGACTCAGCCTATCACAAGGAATCATCCCAAGCTCCTGGACTAAAATCTTA
 ATCATTCTTACCTTAAAGCTTATTACTTGGACCAATTAACCAGCGGCCCAATTTGNAAA
 GGNTATTCTTGAACAGGCTATATGTGTACCGCGCNNTAATGTGTNAATGTATATCCCCAC
 AAACGTGTATGGATTTCTCC**CAGGCGT**CGA

Figure D.11 Nucleotide and deduced amino acid sequences of clone 11. Bold letters indicate the regions of primers.

ATTGGGCGATGGAGGCGAAGATGTAATTTCCAATTTCCATCTATTGTTTGGGAATAAAGCCCA
 TCAAGATTGAATTGAAATTTTTGATAATTTGACTTTGATTTAGATGACTCCCAGTCTGTTTG
 TTTATACCGGAAAGATCGTCTTTCTTTGTTTAGTATTGGTGATGTAAAATTTTAATTATTG
 TGGGTATGTGGTCTGAACCAACGTTCTGACCGGTTTGTAGGTGTGTGGTTCCAGAGAACG
 AGAT**CAGGGCGT**CG

Figure D.12 Nucleotide and deduced amino acid sequences of clone 12. Bold letters indicate the regions of primers.

ATTGGGCGATGTATGTGCAGGTGTTGATTCTTTTGAGTCTTTTATCTTGCCTTATTTTACT
TCTACGTTTTCTTGAGTTCCCCGTTTTTCCAGTTTTCCGATTCTTTTTTCTTGATTTCCCTG
CTATCTGTTTTACTCATTTTAGTGTTTTATTACAGATGTTTTAATCTTAACTAGTATCTGTT
TTCTTGTGGTTTAACATATTAGAACCTGAAGAGTTAGACACGGGGACACAGTCACTTTAACG
AATCTAGGGTTGGCAGTACTGCAGCACTATACCCTAACCCCCCTCCCCCACCCTAGCC**AG**
GGCGTCG

Figure D.13 Nucleotide and deduced amino acid sequences of clone 13. Bold letters indicate the regions of primers.

ATTGGGCGATCTAACCCCTGCCAATTCCTATCTGGGAACTGACTTCTAATCTTCCTAGTTAAG
TACGACTGTCACAAACAAGGCCGCTTGACCTATTCCAGGCAGGACGAACAGTTGAATATAGG
TATACTCCCCTGCAGATTGTTGTATTATGTGGATCTAATGGCAATTTTGCAACAAATTACA
ACTCAGTACAGCTCTCTACAGACCACCTTCCATCGACAGCGTATCTTTAGTGTTGACGCATC
CCCTCGACGC

Figure D.14 Nucleotide and deduced amino acid sequences of clone 14. Bold letters indicate the regions of primers.

AATTGGNGCGGATATNATTCTATAACAATCTCTTTCCCCACAACACGCTCTAGAGATACGCCC
ATATAGTGACACAATATCACTTACCACTAAATATATTTTTTTTTGCAAGGCCATTTTCAAGGT
TAATCCCAAACCATTCACCACCCAACCCATGAGAAATCAACCACCAAACATTATCACTAA
CCACAAAGACCACTTATTGTATGAGACACCACCTCAGGCACACATGCACGGTGAATCAGTG
GTACGACCTGTGTTCTCGCTGAGCCCGAACGACCTAACTGCC**CTCGACGC**

Figure D.15 Nucleotide and deduced amino acid sequences of clone 15. Bold letters indicate the regions of primers.

ATTGGGCGATGTCAATTTACACGTGAATATGTAAGTGTATTTGGAATTATTCATCCCAGGGTA
 ACACTTATTAATTTACAATGATGTAGTTTTTCGACGCGTTACTCATTACCGAGTATAACCATG
 CTATCAAATCTACATTTGGCTAATACCTGTACTGAGCTTTATTCCTGCCCTAAGAAGTTG
 CTTTTCTGATTGTTTTATAACCGCTTGGCTTGATAATATTGTGCTTGCTTGAGAGATCAGT
 CTAGATTTCTCTAATATTATCATTGTACCGTATTATGATCTGGGCCTCGGTTTCAGTTAGTA
TCGCCCAAT

Figure D.16 Nucleotide and deduced amino acid sequences of clone 16. Bold letters indicate the regions of primers.

ATTGGGCGATACTAACTGAACCGAGGCTAGATCATAATACGGTACAAATGATAATATTAGAG
 AAATCTAGGTTGATCTCTCAAGCAAGCACAATATTATCAGGCCAAGCGGTTATAAAACAATC
 CGGAAAAGCAACTTCTTAGGGCAGGAATAAAGCTCAGTACAGGTATTAGCCAAATGTAGAT
 TTTGATAACATGGTATACTCGGTAATGAGTAACGCGTCGAAAACACTACATCATTGTGAAATTA
 ATAATTGTTACCCTGGGATGAATAATTCCAATACACTTACATATTCACGTGTAAATGACAT
CGGCCAAT

Figure D.17 Nucleotide and deduced amino acid sequences of clone 17. Bold letters indicate the regions of primers.

ATTGGGCGATGTCAATTTACACGTGAATATGTAAGTGTATTTGGAATTATTCATCCCAGGGTA
 ACAATTATTAATTTACAATGATGTAGTTTTTCGACGCGTTACTCATTACCGAGTATAACCATG
 TTATCAAATCTACATTTGGCTAATACCTGTACTGAGCTTTATTCCTGCCCTAAGAAGTTG
 CTTTTCCGGATTGTTTTATAACCGCTTGGCCTGATAATATTGTGCTTGCTTGAGAGATCAAT
 CTAGATTTCTCTAATATTATCATTGTACCGTATTATGATCTAGCCTCGGTTTCAGTTAGTAT
CGGCCAAT

Figure D.18 Nucleotide and deduced amino acid sequences of clone 19. Bold letters indicate the regions of primers.

ATTGGGCGATGCAGGCGAAGATGTAATTCCAATGTCCTTCTATTGTTTGGGAATCAAGCCCT
TCAAGACTAAATTCAAAGTTATGATTATCTAATTTAGATTTAAACGACTCCCAGTCTGCTTG
TTTATACCGGAAAGATCGTCTTTCTTTGTTTACTATTGGTGATGTGGAAATCTATAATATGA
TAGGTATGTGGTCTGAACCAACGTTCCGACCGGGTTGTAGGTGTGTGTGATAGGCAAGTGT
GCTCTGAT**CAGGGCGTCG**

Figure D.19 Nucleotide and deduced amino acid sequences of clone 20. Bold letters indicate the regions of primers.

ATTGGGCGATGAAAGTTCATTGGCACGGTGAGCAGCATAATACACTTGCATGGCCAGACTG
TTTGAGTCACAGTCCACAGTCCCACAGACTTTATTCTGATCATTAAAGTTACTAAAATTGAG
ACCTCCCTTAAGGACAAAGTATACTGGAGGTCCAACACTTAGGTACTTTCCTAGATACTCAA
AATATTTTTGTAGGTACGAGTCTTCTGGCATAGACAATTCCTTGATCAAGACCTATGTCAATC
ATTGGCAGAACAGCAATGCTCGAGAACAGCCATCCTACAAACAAGACCACCACAACTGGTCT
GGCAATGCGCGACAAAAGCATGGGAGCATAGACGAGTTTGAAGAATTTCTGTATGGGGCCCT
CAGGTGACGTCCCTTCCTTGTTTGATCCCT**CGACGC**

Figure D.20 Nucleotide and deduced amino acid sequences of clone 21. Bold letters indicate the regions of primers.

ATTGGGCGATCTAACCCCTGCCAATTCCCATCTGGGAACTGACTTCTAATCTTCCTAGTTAAG
TACGACAGTCACAAACAAGGCCGCTTGACCTATTCCAGGCAGGACGAACAGTTGAATATAGG
TATACTCCCGCTGCAGATTGTTGTATTATGTGGATCTAATGGCAATTTTGAACAAATTACA
ACTCAGTACAGCTCTCTACAGACCACCTTCCATCGACAGCGTATCTTTAGTGTTGACGCATC
CCCT**CGACGC**

Figure D.21 Nucleotide and deduced amino acid sequences of clone 22. Bold letters indicate the regions of primers.

AGGCCGCTTAGAAAAGTGAAACAACAAAGCAGACAGTCAACATTCTCAGCCGTTTCGCGCTTC
 AAGACATGTAGATTTTCAGTAGCTCCTTGTAATTGAGGTACGATTCACCTCCTTGCATGTCGT
 TTACTCCTCGTTTAACTATTAATACGTTTTGTGTAATAAATAGCTAGTTCCTACTTCCAT
 GTTTTGCTTTCTACATTGTTTTAATGTGAATAACTAGTTACTATCGTTTGGCTTTGTCTAGTG
 AATAACTTGTAATACTTTGATTTGTTTTGTTTTCTAAACTGTTTTTGCTTTGAAGTAAATA
 ACTAGTTACTCCTTCGAATAGGACCTTAAACAAACACATTTTTACTCTGCTGTCTCCAGGA
 TTATATCCGTT**AAAAGCCCCA**

Figure D.22 Nucleotide and deduced amino acid sequences of clone 24. Bold letters indicate the regions of primers.

AACGGGCAGCCGACGTGGTAGCGCTGTTGGTGTGGTGTGAAGGGTCAGTCATCATATGTT
 TGCTGGAGTCAGGGATTCAGTCCACTATCCGAAATGCAGTCTATGGGGCGCTTGCTGGTGA
 TGGCGAGAGGGAAGTCAGTTGAGTGTAATGCGTCAGCTTCGTTCCGCAATGCTCGATGCTGG
 TGTGGGACAAGCGTAGCCAGTGATCCATAGTCGGGTAATCATCAACGTCATCTTCGCTGAT
 CGGCAAACCGTCTCGGCATCAACAAATTTGAAGGTCCTGAGTAATCCATACCTCCCCCTCT
 GCAGCGCGGAGGGGAGGCACTGACGTACGAGCGGGTATGCAATGACGGGCAAGCACGTT
 ACGCCAGCTCACTGCGCATGCGCAGGAGCGGTAGCCTCGAGCAGGGAAGGCTTGTACCCGA
 CGAGTTTATTTCAAATCTCCAGGCTAGGTCTACACCCTCTTCCTGGGGCACTAATACGTTCT
 CTGGGTCTTACTAGTATCCTGTGGAACGAATAGTT**GCTGCCCGTT**

Figure D.23 Nucleotide and deduced amino acid sequences of clone 28. Bold letters indicate the regions of primers.

ATTGGGCGATACACCGCATGCATAATAAAATGACGTGGGATCCGACTGTAGCAAACAGATAC
 GGATTTGTCAATAAAACGTTAAAACCCGAAATCCACGATGAAGTGTATAATGTAGTACCATC
 ATCGGGGGGAAGGGATGGGGAAATATTTCAATCCATACAATATGACTTTTTGTATGGAATAA
 TCACAAAACAATATCTAAAAAATCTACGCTCTGATTTCAAAGCACGTGATTACGAAACCCAA
 CATAATCCGACCTGAAATTGAGTGGGATGTTGATGTACGATATTTTCGCCTATACCTCACAT
CGCCCAAT

Figure D.24 Nucleotide and deduced amino acid sequences of clone 29. Bold letters indicate the regions of primers.

ATTGGGCGATGTCAATTTACACGTGAATATGTAAGTGTATTTGGAATTATTCATCCCAGGGTA
 AACTTATTAATTTACAATGATGTAGTTTTTCGACGCGTTACTCATTACCGAGTATAACCATG
 TTATCAAATCTACATTTGGCTAATACCTGTACTGAGCTTTATTCTGCCCCAAGAAGTTG
 CTTTTCTGATTGTTTTATAACCGCTTGGCCTGATAATATTGTGCTTGCTTGAGAGATCAAT
 CTAGATTTCTCTAATATTATCATTGTACCGTATTATGATCTGGCCTCGGTTCAAGTTAGTAT
CGCCCAAT

Figure D.25 Nucleotide and deduced amino acid sequences of clone 30. Bold letters indicate the regions of primers.

ACTAGTGATTGCGTCGAGGGCAATTAACGACAAGCTGTGAGGGCATCCAGGTTGTTGCAGT
 TTTCTGTTGCAAGTTTTCTAGCAAGAGATGCAATGGCTTTGCAAAGGTCATTTGCAGCTGAC
 CCAAAGTTGCTGCTGCTCAGGATGGTTTTCCATGCCTTTGCGTCGAGTCCAGAAGGTCCAGC
 TGCACCATGTGTATGCAATGCATGTGTTCTTATCTTTTCGCCGG

Figure D.26 Nucleotide and deduced amino acid sequences of clone 52. Bold letters indicate the regions of primers.

ACTAGTGATTTATGGGCGATACACCGCATGCATAATAAAATGACGTGGGATCCGACTGTAGC
 AAACAGATACGGATTTGTCAATAAAACGTTGAAACCCGATATCCACGATGAAGTGTATAATG
 TAGTACCATCATCGGGGGGAAGGGACGGGGAAATATTTCAATCCATACAATATGACTTTTTG
 TATGGAATAATCACAAAACAATATCTAGAAAATCTACGCTCTGATTTCAAAGCACGTGATTA
 CGAAACCCAACATAATCCGACCTGAAATTGAGTGGGATGTTGANTGTACGATATTGTCGCCT
 NTACCTCACAGTCGCCCAATAATCG

Figure D.27 Nucleotide and deduced amino acid sequences of clone 58. Bold letters indicate the regions of primers.

ACTAGTGATTTATGGGCGATAACATATGATACCAAATCTTAAGACATCAACAACAATAATAA
 AGATAATAGCCAGTGATCCATAGTCGGGTAATCATCAACGTCATCTTCGCTGATCGGCAAAA
 CCGTCTCGGCATCAACAATTTGAAGTCTCTGAGTAATCCATACCTCCCCCTCTGCAGCGCG
 CGAGGGGAGGCACTGACGTCACGAGCGGGTATGCAATGACGGCAAGCACGTTACGCCAG
 CTCACTGCGCATGCGCAGGAGCGTAATAATTAATGTTGCTGCTGATGATTACAATGATATTG
 ATAATGATATCAACAACGGTAATAGTAA

Figure D.28 Nucleotide and deduced amino acid sequences of clone 62. Bold letters indicate the regions of primers.

Penaeus monodon clone TUZX4-6:86 microsatellite sequence

Length = 517

Score = 149 bits (75)

Expect = 3e-33

Identities = 90/95 (94%)

Strand = Plus / Plus

```

Query: 171   ttttaacggtaggttcatgtctgagccaccgtagtcacagtatgataacttaattgtcgg 230
              ||| | | | | | | | | | | | | | | | | | | | | | | | | | | | | | |
Sbjct: 168   ttttaacggtaggttcatgtctgagccgccggtggtcacagcatgataacttaattgtag 227

Query: 231   ttcatgttgatgctcctggagtgagtacgtgg 265
              ||| | | | | | | | | | | | | | | | | | | | | | | |
Sbjct: 228   ttcatgttgatgctcctggagtgagtacgtgg 262
  
```

Figure D.29 The results of blast in GENBANK database and the high significant alignment of a deduced amino acid sequence of clone 4. The website (<http://www.ncbi.nlm.nih.gov/blast>) was used in this analysis.

Penaeus (Litopenaeus) vannamei microsatellite TUMXLv10.221 sequence

Length = 600

Score = 93.7 bits (47)

Expect = 3e-16

Identities = 83/95 (87%)

Strand = Plus / Plus

```

Query: 196   ttttaagttgtacctattgtttagttttattggttactgtagctactccctcgcttacacaa 255
              ||| | | | | | | | | | | | | | | | | | | | | | | | | | |
Sbjct: 235   ttttaagttgtacctattgtttagttttattattacagaagccactctttcgcttacacta 294

Query: 256   cagaattccacgatattcttttctaggcggttgg 290
              ||| | | | | | | | | | | | | | | |
Sbjct: 295   tagaattctacgatgttcttttctaagagtttgg 329
  
```

Figure D.30 The results of blast in GENBANK database and the high significant alignment of a deduced amino acid sequence of clone 9. The website (<http://www.ncbi.nlm.nih.gov/blast>) was used in this analysis.

ENSANGP00000010415 [*Anopheles gambiae* str. PEST]

Length = 1548

Score = 146 bits (369)

Expect = 1e-34

Identities = 68/125 (54%)

Positives = 91/125 (72%) Frame = -2

```

Query: 375  GTSPEGPIQKFFKLVYAPMLLSRIARPVVVVLVFGWLFSSIAVLP MIDIGLDQELSM PED196
          G  EG + KFFK +Y P+++ R      V+++F GWL SSI AV P IDIGLDQELSM P D
Sbjct: 776  GNIGEGLLYKFFKSIYVPFVMKRPVRVAVMIVFFGWLCSSIAVAPHIDIGLDQELSM PGD 835

Query: 195  SYLQKYFEYLGKYL SVGPPVYFVLKGG LNFSNLNDQNKVC GTVDCDSNSLAMQVYAAHR 16
          S++ KYF YL +YLS+GPPVYFV+K GLN+S +NDQN +CG C ++SL+ Q+Y A++
Sbjct: 836  SFVLKYFRYLQQYLSIGPPVYFVVKGNLNYSTMNDQNLICGGQYCNLDSLSTQLYIASKQ 895

Query: 15   ANRTF 1
          T+
Sbjct: 896  PQSTY 900

```

Niemann-Pick type C1 disease protein [*Oryctolagus cuniculus*]

Length = 1286

Score = 128 bits (322)

Expect = 3e-29

Identities = 63/120 (52%)

Positives = 85/120 (70%)

Frame = -2

```

Query: 363  EGPIQKFFKLVYAPMLLSRIARPVVVVLVFGWLFSSIAVLP MIDIGLDQELSM PEDSYLQ 184
          E  ++FFK Y+P+LL  RP+V ++FVG L  SIAVL ++IGLDQ LSMP+DSY+
Sbjct: 823  ESYLFRFFKNSYSP LLLKDWMP IVI AVFVGVLSFSIAVLNKVEIGLDQSLSM PDDSYVV 882

Query: 183  YFEYLGKYL SVGPPVYFVLKGG LNFSNLNDQNKVC GTVDCDSNSLAMQVYAAHRANRT 4
          YF+ LG+YL  GPPVYFVL+ G N+++L QN VCG +CD++SL Q++ AA  N T
Sbjct: 883  DYFKSLGQYLHAGPPVYFVLEEGHNYTSLQGQNMVCGGLGCDNDSL VQQIFNAAQLDNYT 942

```

Figure D.31 The results of BLAST from GENBANK database and the high significant alignment of a deduced amino acid sequence of clone 21 (<http://www.ncbi.nlm.nih.gov/blast>)

APPENDIX E

Table E.1 Protein concentration in haemolymph thermal treatment for 6 h

(hours)	15°C	Control (27°C)	30°C	33°C	35°C
0	13.95	50.00	80.60	53.40	73.40
	20.91	47.13	52.20	77.40	68.40
	12.95	41.62	63.80	64.20	53.94
	17.33	49.25	70.60	93.60	99.60
	13.56	39.25	64.80	76.20	79.80
3	28.81	39.69	66.40	72.96	75.03
	6.34	49.66	61.40	93.00	59.80
	12.25	42.75	69.60	54.20	78.20
	24.26	46.63	91.80	54.60	85.40
	9.42	39.34	53.60	90.40	72.20
6	24.56	48.69	64.68	70.64	72.88
	13.05	43.51	88.80	66.20	54.60
	23.35	35.63	61.60	52.60	62.20
	9.42	40.29	62.60	70.20	54.80
	23.15	46.43	58.60	69.20	98.40
12	21.03	41.48	60.48	63.76	66.12
	16.87	47.26	68.40	67.00	62.20
	17.65	37.55	79.00	72.60	56.20
	18.00	44.26	62.80	69.40	70.20
	30.42	39.96	71.40	81.40	93.80
24	20.23	38.98	68.12	68.64	69.00
	25.17	42.20	60.60	75.20	82.20
	27.78	39.32	59.40	83.40	41.80
	36.84	42.41	73.20	48.80	57.80
	13.86	39.77	58.60	69.80	89.80
72	26.94	44.03	66.32	65.88	66.60
	22.44	43.88	51.60	71.00	55.80
	31.18	33.26	52.00	67.00	86.20
	20.22	38.66	76.20	48.60	62.20
	29.31	37.24	72.60	54.30	45.80

Table E.2 Glucose concentration in haemolymph thermal treatment for 6 h

(hours)	15°C	Control (27°C)	30°C	33°C	35°C
0	46.85	45.12	35.24	35.21	25.65
	20.91	27.5	25.69	30.15	30.33
	33.43	30.58	50.01	35.4	20.56
	19.12	40.01	20.69	40.26	37.51
	23.59	25.67	30.14	30.58	55.01
3	23.59	35.03	160.14	70.44	125.03
	34.33	22.51	30.99	75.65	55.26
	24.49	30.98	60.14	60.11	60.45
	37.91	40.01	30.59	45.63	45.7
	12.86	32.56	20.98	70.32	90.11
6	19.12	35.22	35.47	115.01	90.2
	23.59	30.54	35.67	75.34	90.47
	27.17	40.11	30.12	95.74	85.96
	100.52	40.68	25.26	65.11	105.51
	19.12	25.12	10.25	80.02	85.03
12	21.80	20.98	95.02	90.17	100.01
	19.12	25.47	120.14	100.03	80.19
	24.49	60.05	65.22	70.68	115.32
	22.70	30.24	40.8	90.03	105.12
	24.49	50.11	55.48	95.05	90.94
24	45.06	35.48	150.11	85.44	95.21
	40.59	30.29	65.28	90.04	100.34
	47.74	17.56	85.67	70.55	7.5.55
	43.27	45.13	5.97	60.33	85.64
	30.75	40.65	30.56	95.84	80.88
72	43.27	50.14	65.15	45.62	40.43
	45.06	20.69	40.55	60.25	60.51
	23.59	30.54	60.01	50.55	50.02
	36.12	30.68	37.53	35.44	45.06
	11.07	40.16	60.33	55.28	90.81

Table E.3 Protein concentration of the haemolymph from heat-induced shrimps after *vibrio* exposure

hours	sample	control	heat induced	un-induced
0	1	46.6	63.3	49.1
	2	70	75.9	49.3
	3	44.6	60.2	42.4
	4	62.3	73.2	35.7
	5	32.9	64.3	64.2
	6	52.4	57.4	66.2
3	1	46.6	73.2	50.1
	2	33.7	67.4	60.5
	3	60.5	64.2	52.6
	4	46.7	71.9	44.2
	5	66.9	61.7	67.8
	6	64.7	71.7	47.6
6	1	51.5	74.2	52.6
	2	46.5	69.6	66.9
	3	60.9	58.8	53.9
	4	44.7	76.9	47.9
	5	71.1	88.7	67.8
	6	42.5	67.2	52.3
12	1	22.9	82.9	69.9
	2	64.9	72.9	68.4
	3	61.5	76.1	70.5
	4	52.8	64.6	52.7
	5	52.2	69.7	67.9
	6	50.1	52.1	47.8
24	1	59.1	73.5	42.5
	2	60.8	60.3	43.9
	3	30.6	71.7	68
	4	50.9	48.2	52.9
	5	58.1	74.5	53.7
	6	59.1	76.5	64.6
72	1	32.7	72.7	63.4
	2	47.2	50.5	49.5
	3	62.1	61.4	51.9
	4	55.4	56.7	62
	5	62.9	71.1	31.9
	6	42.7	64.9	53.1
120	1	36.1	58.8	63.8
	2	62.8	45.4	47.9
	3	53.2	66.7	56.2
	4	67.6	60.9	45.4
	5	41.3	64.7	42.1
	6	58.1	52.6	60.8
168	1	60.4	73.4	58.2
	2	73.9	35.5	61.6
	3	49.31	55.3	54.3
	4	36.3	34.2	30.6
	5	47.9	65.8	61.2
	6	40.9	43.3	52.6



Table E.4 Glucose concentration of the haemolymph from heat-induced shrimps after *vibrio* exposure.

hours	sample	control	heat induced	un-induced
0	1	32.54	43.27	40.59
	2	40.59	45.06	49.53
	3	37.91	44.17	34.33
	4	27.17	43.27	48.64
	5	38.80	54.01	42.38
	6	39.69	32.54	36.12
3	1	46.85	37.91	39.69
	2	25.38	46.85	42.38
	3	32.54	56.69	34.33
	4	42.38	59.37	51.32
	5	26.28	46.85	38.80
	6	39.69	55.80	37.01
6	1	35.22	53.11	40.59
	2	33.43	58.48	35.22
	3	50.43	46.85	34.33
	4	34.33	61.16	31.64
	5	41.48	62.06	50.43
	6	27.17	62.95	34.33
12	1	31.64	63.85	33.43
	2	48.64	53.11	39.69
	3	30.75	57.58	26.28
	4	41.48	53.11	39.69
	5	30.75	73.69	45.96
	6	33.43	66.53	47.74
24	1	46.85	48.64	38.80
	2	34.33	43.27	34.33
	3	29.85	49.53	32.54
	4	28.96	52.22	45.06
	5	35.22	55.80	43.27
	6	37.01	65.64	29.85
72	1	33.43	51.32	42.38
	2	35.22	48.64	38.80
	3	41.48	42.38	29.85
	4	33.43	37.91	47.74
	5	29.85	50.43	37.01
	6	52.22	45.06	33.43
120	1	28.07	28.96	43.27
	2	45.06	43.27	48.64
	3	45.06	31.64	41.48
	4	28.07	53.11	28.96
	5	36.12	38.80	31.64
	6	25.38	28.07	34.33
168	1	35.22	37.01	43.27
	2	25.38	33.43	34.33
	3	41.48	36.12	29.85
	4	37.91	41.48	38.80
	5	46.85	42.38	37.91
	6	24.49	28.96	32.54

BIOGRAPHY

Miss Kanchana Doungpunta was born on July 23, 1978 in Rayong. She graduated with the Bachelor Degree of Science in Department of Biology, Faculty of Science, Burapha University in 2000. She has been a graduate student in the Master's Degree in Biotechnology program, Faculty of Science, Chulalongkorn University since 2004.

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