CHAPTER II

HISTORICAL

1 Distribution

1.1 Chemical and botanical aspects of isoquinoline alkaloids

A precise definition of the term "alkaloid" (alkalilike) is some what difficult because there is no clear-cut boundary between alkaloids and naturally occuring complex amines. Typical alkaloids are derived from plant sources, they are basic, they contain one or more nitrogen atoms (usually in a heterocyclic ring) and they usually have a marked physiological action on man or other animals. The nitrogen of alkaloids, taken in their broadest sense, may have a nitrogen atom which is primary (mescaline), secondary (ephedrine), tertiary (atropine) or quaternary (one of the N-atoms of tubocuraline), and this factor affects the derivatives of the alkaloid which can be prepared and the isolation proceduces. In the plant alkaloids may exist in the free state, as salts or as amine or alkaloid N-oxides. But, for practical considerations it must be more restrictive with some exceptation. There are numerous groups of alkaloids isolated from the lower plants, animals, microorganism, flowering plants and marine natural products.

In the higher plant system of Engler, there are 60 orders, and 34 of these contain alkaloid bearing species. The most important alkaloid - containing families are the Liliaceae, Amaryllidaceae, Compositae, Lauraceae, Ranunculaceae, Menispermaceae, Papaveraceae, Leguminosae, Rutaceae, Loganiaceae, Apocynaceae, Solanaceae and Rubiaceae

Among those alkaloids, isoquinoline alkaloids have played an important part in the development of the chemical and biological sciences.

Alkaloids with the isoquinoline ring system or those derived from a phenylalanine unit and therefore related structurally and biogenetically to the isoquinolines comprise at least 25 different types as follow (Govindachari and Viswanathan, 1972)

Type of isoquinoline alkaloids

- 1 Simple isoquinolines
- 2 1-phenylisoquinolines
- 3 1-benzylisoquinolines
- 4 Cularine group
- 5 Phthalideisoquinolines
- 6 Protoberberines
- 7 Protopine group
- 8 Benzophenanthridines
- 9 Aporphines
- 10 Proaporphines
- 11 Dibenzo pyrrocolines
- 12 Morphine group
- 13 Protostephanine

- 14 Hasubanans
- 15 Pavine group
- 16 Ochotensine group
- 17 Rhoeadine group
- 18 Bisbenzyl isoquinolines
- 19 Emetine group
- 20 Erythrinans
- 21 Amaryllidaceae
- 22 Indoloisoquinolines
- 23 1-Phenethylisoquinolines
- 24 Isopavine group
- 25 Terpene alkaloids

The number of isoquinoline alkaloids of known structure is approximately to 1,200 both in tetrahydroisoquinoline and quaternary isoquinoline salts. The majority of isoquinoline alkaloids have been isolated from the nine plant families such as Anonaceae, Berberidaceae, Fumariaceae, Hernandiaceae, Lauraceae, Menispermaceae, Papaveraceae, Ranunculaceae and Rutaceae. But they are also known to occcur in other plant families, e.g. Alangiaceae, Amaryllidaceae, Cactaceae, Combretaceae, Convolvulaceae, Euphorbiaceae, Leguminoseae, Magnoliaceae, Monimiaceae, Nymphaeaceae and Rubiaceae. With respect to their structural features, the isoquinoline alkaloids can be devided into two main classes. The first class is that of the simple isoquinoline alkaloids. The simple isoquinolines are structurally the simplest of the isoquinoline alkaloids.

They are usually bicyclic, although tricyclic species such as peyoglutan and mescalotam are also included among them. The nitrogen function in ring B is often tertiary and N-methylated, but it may also be secondary, N-formylated, N-acetylated, N-ethylated or oxidized to the imine stage. Quaternary simple isoquinoline, e.g. lophotine and 2-methyl-6,7-dimethoxy isoquinolinium salt, have also been isolated. Of more than passing interest is pilocereine, the only trimeric isoquinoline alkaloid fully characterized. Simple isoquinolines display great variety in their substitution pattern, depending, of course, upon their biogenetic origin. Most simple isoquinolines have been obtained from the Cactaceae, but they also occur among the Alangiaceae, Annonaceae, Berberidaceae, Euphorbiaceae, Leguminosae, Menispermaceae, Papaveraceae and Ranunculaceae (Menacherry et al, 1986).

The second class is the benzyl-derived isoquinoline alkaloids. They do not present a structural uniformity having benzylisoquinolines act as precursors to so many other naturally occurring isoquinoline type e.g. isoquinolines, pavines, isopavins, bisbenzylisoquinoline, cularines, protoberberines, erythrina base and others. Because relatively a large number of isoquinoline alkaloids in this class are known, their occurrence are distributed in many plant families.

1.2 Botanical aspects of the Menispermaceae

The Menispermaceae is one of a large family containing approximately 73 genera and about 350 species, wihch are almost entirely tropical. The exceptions being Menisperum, a northern temperate genus with 2 disjunct species in North America and Northern Asia, and a few species of Cocculus which extend into North America and temperate Asia.

There are 30 genera of this family occur in Asia, 30 in Africa, 22 in America and 10 in Australia to the Pacific. Of the 25 Malesian genera 20 occur in continental Asia, and 6 occur in Africa of which 2 (Cissampelos and Cocculus) are also in America. Of the Malesian genera 9 are shared with Australia and of these 6 extend into Asia, Legnephora is limited to central and East Malesia, Corrania and Sarcopetalum occur in New Guinea. Only 2 of the Malesian genera are endermic, Chaenandra and Macrococculus, both in New-Guinea. The Menispermaceae is characterized by dioecious woody or sometimes herbaceous climbers, rarely erect shrubs or trees (Cocculus sp.); tubers sometimes present (Stephania spp.); sometimes producing exudate or rarely latex (Fibraurea; Tinomiscium). Wood often with concentric rings or arcs of vascular bundles separated radially by interfascicular rays, or vascular bundles in one ring, wood sometimes yellow. Young shoots often tendrilliform. Young stems usually drying longitudinally striate. Stipules absent. Leave spiral, simple, often palmatinerved at base and sometime peltate, or pinninerved, margin

usually entire, sometimes boardly crenate, sometimes deeply 3-5 lobed, petiole often swollen at base, sometimes also at apex, sometimes leaving a raised discoid scar on the stem.

Inflorescences axillary or on defoliate branches or cauliflorous, solitary or fasciculate, various in form, often cymes, thyrses or pseudoracemes, branching of cymes rarely umbelliform (Stephania spp.), flowers rarely in a disciform capitulum (Stephania spp.); female usually fewer flowered than male, female rarely with accrescent bracts (Cissampelos spp.). Flowers small, usually green, yellow or white, actinomorphic or female sometimes zygomorphic. Sepals usually in 1-2 (-4) whorls of 3, or 1 whorl of 4, the outer whorl(s) smallest, imbricate but the innermost whorl sometimes valvate and sometimes connate, sepals rarely spirally arranged (Hypserpa); in female sometimes reduced to 1 or 2. Petals mostly 3-6 in 1 or 2 whorls or 0, free or sometimes + connate, usually smaller than the sepals, rarely larger (Sarcopetalum), the lateral edges or lobes often inflexed and sometimes clasping the opposite stamen, often glandular within, in female sometimes reduced to 1 or 2.

Stamens mostly 3 or 6, sometimes 9 or up to about 40, often free and opposite a petal, or variously connate, sometimes forming a peltate synandrium, connective sometimes adaxially or abaxially thickened, rarely terminally prolonged (Macrococculus); anthers introrse to extrorse with dehiscence longitudinal to transverse. Staminodes sometimes present in female, usually subulate, carpels free,

usually 3 or 6, sometimes 1 or to 12, sometimes borne an a short gynophore; style terminal when present; stigma often sessile, reflexed and lobed or divided. Pistillodes 0 in male.

Ovules 2 reducing to 1 in development, attached ventrally. Fruits of 1-6 (-10) drupes sometimes borne on an enlarged globose, discoid or columnar carpophore which is rarely shortly branched (Anamirta, Tiliacora).

Drupes sometimes narrowed at base into a stipe, style-scar terminal, ventral or close to base; exocarp membranous to coriaceous, mesocarp fleshy; endocarp usually bony, rarely papyraceous to crustaceous (*Pycnarrhena* spp.) rugose, tuberculate, spiny, ridged or variously ornamented on at least the dorsal surface, sometimes smooth or surface fibrous, usually with a condyle; i.e. a ventral sometimes hollow intrusion into the seed cavity around which the seed is curved, or a ventral groove, cavity or chamber; the condyle when hollow often 2-chambered and with 2 lateral or ventral apertures or condyle septiform or lamelliform, then sometimes centrally perforate.

Seed often horseshoe-shaped; endosperm present or absent, sometimes ruminate. Embryo usually either elongate and with semiterete or flattened contiguous cotyledons or flat and very thin with divaricate foliaceous cotyledons, sometimes broadly ellipsoidal with thick contiguous cotyledons, rarely cotyledons much folded (Arcangelisia); radicle vary small. (Forman, 1986)

According to Forman the members of Menispermaceae were divided into 5 tribes, there are Coscinieae, Menispermeae, Tiliacoreae, Tinosporeae and Fibraureeae. But two of these, Fibraureeae and Tinosporeae should probably be combined. The tribes in Asia are characterized by the following combinations of characters.

Coscinieae: sepals imbricates. Petals O, stamens either all or only the inner 3 connate. Carpels 3-6. Drupe with style-scar sublateral towards base on lateral. Endocarp smooth or fibrillo-pilose, subglobose with condyle obsolete, or subhemispherical with condyle deeply intrusive and 2-chambered. Endosperm present, sometimes ruminate. Seed broadly ellipsoidal or cup-shaped. Embryo with thin foliaceous divaricate cotyledons which are sometimes much folded.

Menispermeae: sepals usually free in 1-few whorls or sometimes connate when in 1 whorl, the innermost whorl sometimes valvate, or sepals spiral. Petals (0-) 3-6 (-9) sometimes connate. Female flowers with perianth sometimes reduced to 1-2 parts. Stamens free or partly connate or united into a peltate synandrium. Carpels 1-6. Drupe strongly curved with style near base. Endocarp with thorseshoe-shaped dorsal region usually ornamented with projections or transverse ridges; condyle deeply intrusive, either lamelliform and the obovate with the seed-cavity curved around its margin or hollow with 1-2 chambers, sometimes perforate. Endosperm usually present, but absent in Pachygone. Seed elongate, strongly curved. Embryo elongate

and curved with narrow contiguous cotyledons.

Tiliacoreae: sepals imbricate or inner whorl valvate and sometimes connate. Petals rarely absent. Stamens free or connate. Carpels 3-10. Drupe with style-scar near base or lateral. Endocarp smooth, wrinkled, rugose or coarsely reticulate; straight and condyle absent or curved with condyle intrusive and septiform. Endosperm usually absent, but present and ruminate in Tiliacora. Seed ellipsoidal, straight. Embryo with thick accumbent cotyledons or elongate and strongly curved with elongate contiguous cotyledons.

Tinosporeae (include Fibraureeae): sepals imbricate, rarely connate at the base. Petals 6 or 0. Stamens free or united into a peltate synandrium. Carpels 3 (-4). Drupe with style-scar terminal. Endocarp spiny, verrucose, rugose or smoth, condyle a ventral hollow or longitudinal groove or deeply intrusive and clavate endosperm present, sometimes ventrally ruminate. Seed usually straight and ventrally hollowed or grooved, sometimes cup-shaped. Embryo with foliaceous divericate or imbricate cotyledons.

There are 22 genera, and 51 species in Thailand, of with 9 species are endermic (Shown by asterich). All of them are as follows:

- 1 Albertisia papuana Becc.
 Albertisia puberula Forman.*
- 2 Anamirta cocculus (Linn.) Wight & Arn

[Waidin (หวายดิน), kho khlan (โคคลาน) (Central) ; thao kha nom (เถาบะโนม), lumpri (ลุมพรี) (South-Eastern) ; mae nam nong (แม่น้ำนอง) (Northern) ; thao wan thong

(เถาวัลย์ทอง) (South-Western)]

3 Arcangelisia flava (Linn.) Merr.

[Khamin khruea (ปมั้นเครือ) (South-Eastern) ; khamin ruesi (ปมั้นฤาชี), hap (ฮับ) (Peninsular)]

- 4 Aspidocarya uvifera Hook.J.& Thoms.
- 5 Cissampelos hispida Forman*

Cissampelos pareira Linn. var. hirsuta Forman.

[Khong khamao (บงเบมา) (Northern) ;

khruea manoi (เครื่อหมาน้อย) (Eastern) ; kon pit (กันปิด) (South-Western) ; krung khamao (กรุงเขมา), sifan (สีพัน) (Peninsular)]

6 Cocculus hirsuta (Linn.). Theob.

Cocculus laurifolius DC.

[Yan nanton (ย่านน่านต้น) (North-Eastern, Central); sakae dong (สะแกดง) (North-Eastern) ; suramarit (สุรามฤต) (Eastern)]

Cocculus orbiculatus (Linn.) DC.

7 Coscinium blumeanum Miers

Coscinium fenestratum (Gaertn.) Colebr.

[Khruea hen (เครือเห็น) (North-Eastern);

khamin khruea (ปฏินเครือ) (South-Eastern)]

8 Cyclea atjehensis Forman

Cyclea barbata Miers

[Krung badan (กรุงบาดาล) (South-Eastern) ;

krung khamao (กรุงเขมา) (Peninsular).]

Cyclea laxiflora Miers

Cyclea polypetala Dunn

Cyclea varians Craib*

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- 9 Diploclisia glaucescens (Bl.) Diels
- [Ma nim dam (มะหนิมดำ), duk khruea (ดูกเครือ)
 (Northern) ; khruea sai kai (เครือได้ไก่) (Shan/Northern) ;
 tap tao (ตับเต่า) (Peninsular)]
 - 10 Fibraurea tinctoria Lour.

[Khamin ruesi (ปมิ้นฤาษี) ; khamin khruea (ปมิ้นเครือ) ; man miat (มันเมียด) (Peninsular) ; thaowan thong (เถาวัลย์ทอง) (South-Western); kamphaeng chet chan (กาแพงเจ็ดขั้น) (Central)]

- 11 Haematocarpus validus (Miers) Bakh.f.ex. Forman
- 12 Hyperpa nitida Miers
 [Haen kuem (แฮนก็ม) (North-Eastern)]
- 13 Limacia blumei (Boerl.) Diels

 Limacia oblonga Hook.f. & Thoms.

 Limacia scandens Lour.
- 14 Pachygone dasycarpa Kurz.

[Nam phrom (น้ำพรม) (Northern) ; ya nang chang (หญ้านางช้าง) (Eastern)]

Pachygone odorifera Miers

- 15 Parabaena sagittata Miers
 [Phak nang (ผักหนัง) (Northern)]
- 16 Pericampylus glaucus (Lamk.) Merr.

[Salit hom kha (สลิดหมคา) (Northern) ; yan tap tao (ย่างตับเต่า) (Peninsular)]

- 17 Pycnarrhena lucida (Teijsm. & Binn.) Miq.
 [Ya nang ton (ย่านางตัน) (South-Western)]
 Pycnarrhena poilanei (Gagnep.) Forman
- 18 Sinomenium acutum (Thunb.) Rehder & Wilson
- 19 Stephania brevipes Craib*

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[Bua khruea (บัวเครือ) (Northern)]
          Stephania capitata (Bl.) Spreng.
          Stephania crebra Forman*
          Stephania elegans Hook.f. & Thoms.
          [Se-khi-pho (เสีมิพอ) (Karen/Northern)]
          Stephania glabra (Roxb.) Miers
          [Phanang nang (Wũ V V) (Northern)]
          Stephania glandulifera Miers
          Stephania japonica (Thunb.) Miers
          [Kon pit (กันปิด), bai kon pit (ใบกันปิด), (Central);
pang pon (บังปอน) (Northern); tap tao (ตับเต่า), yan pot
(ย่านปด) (Peninsular)]
          Stephania oblata Craib
          Stephania papillosa Craib*
          Stephania pierrei Diels
          [Bua khruea (บัวเครื่อ) (North-Eastern); bua bok
(บัวบก) (South-Western, Eastern and Central)
kot hua bua (โกฐหัวบัว), sabu lueat (สมู่เลือด) (Central)]
          Stephania reticulata Forman
          [Tap tao (ตับเต่า) (Peninsular)]
          Stephania rotundra Lour.
          Stephania suberosa Forman*
          Bua bok (บัวบก) (Central) ; boraphet phung chang
(บอระเพ็ดพุงช้าง) (South-Western)]
          Stephania subpeltata H.S. Lo.
          Stephania tomentella Forman*
          Stephania venosa (Bl.) Spreng
          [Plao lueat khruea (เปล้าเลือดเครื่อ) (Northern);
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cho koe tho (ชื่อเกอะทอ) (Karen/Northern); krathom lueat (กระท่อมเลือด) (North-Eastern); kling klang dong (กลิ้งกลางดง) (South-Western); boraphet yang daeng (บอระเพ็ดยางแดง) (Peninsular).]

20 Tiliacora triandra (Colebr.) Diels
[Choi nang (จือยนาง) (Northern) ; thao ya nang
(เถาย่านาง) (Eastern, Central) ; thaowan khieo (เถาวัลย์แก้ว)
(South-Eastern)]

21 Tinomiscium petiolare Hook.f. & Thoms.
[Pharai ho thong (ฟ้าร้ายห่อทอง) (Peninsular)]

22 Tinospora baenzigeri Forman
[Chung cha ling (จุงจาลึง) , ching cha chali
(ชิงช้าชาลี) (Central).]

Tinospora crispa (Linn.) Hook.f. & Thoms.
[Boraphet (บอระเพ็ด) (Central)]

Tinospora siamensis Forman*

Tinospora sinensis (Lour.) Merr.

[Ping kaling (ปิงกะลิง) (Northern) ; sali thao chali (สลี เภาชาลี) (Central) ;] (Forman, 1991)

All members of this family seem to produce phenylalanine and tyrosine-derived isoquinoline alkaloids. Aporphines, bisbenzyl-isoquinolines, and quaternary and intensely coloured protoberberines such as berberine and its allies are most typical of the family. In some genera these more usual types of isoquinoline alkaloids are accompanied by less common or even rare types of benzylisoquinoline-related alkaloids. Such types of Menispermaceous alkaloids are the hasubanans, the azafluoranthenes and

related tropolo-isoquinolines, and the dibenzazonines and related Erythrina alkaloids. Moreover, in recent time, pavine-type and aristolactam-type alkaloids were detected in the family. Alkaloid chemistry clearly allocates Menispermaceae to Polycarpicae with the position of one of its more specialized members. Other groups of constituents which seem to be rather characteristic of the family are the bitter and more or less toxic principles, which are sesquiterpenoids like picrotoxin or diterpenoids such as columbin and tinophyllone (Hegnauer, 1969, 1973). It is perhaps not solely accidental that quaternary protoberberine alkaloids like berberine, columbamine, jatrorrhizine and palmatine and diterpinoid bitter principles such as tinophyllone also occur in some genera of Rutaceae. A third group of phytoconstituents, the cyclitols, is known to be accumulated by members of several genera of Menispermaceous plants; it is represented by the deastereoisomeric cyclohexanepentols (+) quercitol and (-) quercitol (viburnitol); they are presently known to occur in the genera Cissampelos, Cocculus, Cyclea, Legnephora, Menispermum, Pachygone, Stephania, Tiliacora and Triclisia. The phenolic constituents were studied only superficially hitherto. Leaves contain flavonols or flavones, or both, but seem to representatives with trihydroxylated B-ring and true tannins. All other classes of phyto-constituents were neglected by phytochemist. Nevertheless, some incidental observations might prove in future to be taxonomically relevant.

1.3 Botanical aspects of the Coscinieae tribe

The tribe Coscinieae is one of the smallest in the Menispermaceae, consisting of only three small genera:

Anamirta, Arcangelisia and Coscinium. The five species which make up the tribe all occur within the Indo-Melesian region, these are:

- 1) Anamirta cocculus (L.) Wight & Arn.
- 2) Arcangelisia flava (L.) Merr.
- 3) Arcangelisia tympanopoda (Lauterb& K.Schum.) Diels
- 4) Coscinium blumeanum Miers
 - 5) Coscinium fenestratum (Gaertn.) Colebr.

In spite of the small size of the tribe, some important morphological feature vary considerably between genera. The leaves are often peltate in Coscinium but not peltate in the other two genera. The inflorescence in Coscinium are composed of peduncled capitula, but they are paniculate in the other genera. In the fruits of Arcangelisia, the condyle is absent or inconspicuous and consequently the seed is broadly ellipsoidal in shape. In Anamirta and Coscinium, however, the condyle is deeply intrusive into the seed-cavity with the seed formed around it, resulting in a deeply concave, subhemispherical seed. In Arcangelisia the seeds are distinctive in having deeply ruminate endosperm. Taking together, the gross morphological characters within the Coscinieae indicate that Anamirta and Arcangelisia are more closely related to each other than either are to Coscinium. (Forman, 1978).

2. Chemistry of the Alkaloids

2.1 Alkaloids isolated from the Menispermaceae

The alkaloids of the Menispermaceae have received considerable attention for a long time. The Menispermaceae are widely distributed in the tropical countries and have been studied throughout the world. The vast majority of alkaloids found in the Menispermaceae are of the benzylisoquinoline type. The alkaloids and their structures which have been reported in 68 species of Menispermaceae are shown in talble 1 & 2.

The structure of alkaloids found to date in the Menispermaceae classified under general structural type and made a precise of type are shown as follows:

Precise
A
Benz
Bis
Н
Pa
P
М
Misc

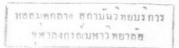


Table 1 Alkaloids isolated from the Menispermaceae (Thonber, 1972)

Plant species	Alkaloids present	Formulae
Albertisia papuana	Aromoline	Bis 62
	Cocsoline	Bis 35
	Daphnoline	Bis 67
	Dehydrotelobine	Bis 40
	Homoaromoline	Bis 68
	Isotrilobine	Bis 36
	Lindoldhamine	Bis 29
	Obaberine	Bis 70
	O-methylcocsoline	Bis 37
	Oxyacanthrine	Bis 71
Anamirta cocculus	Berberine	P2
	Columbamine	P4
	L-8-oxotetrahydropalmatine	P
	Magnoflorine	A7
	Menispermine	-
	Palmatine	P2
	Paramenispermine	-
Arcangelisia flava	Berberine	P1
	Columbamine	P4
	Dehydrocorydalmine	P7
	Homoaromoline	Bis68

		,
	Hydroxyberberine	P
	Jatrorrhizine	P6
	Pycnarrhine	Misc.7
	Shobakunine	_
	Thalifendine	P15
Burasaia madagascariensis	Columbamine	P4
	Jatrorrhizine	P6
	Palmatine	P2
Chasmanthera dependens	Bisnorargemonine	Misc.6
(Ohiri, 1983)	Columbamine	P4
	Coreximine	P19
	Govanine	P18
	Jatrorrhizine	P6
	Magnoflorine	. A7
	Pallidine	м9
	Palmatine	P2
	Pseudocolumbamine	P20
Chondodendron candicans	d-Bebeerine	Bis 19
	Isochondodendrine	Bis 49
Chondodendron limaciifolium	Isochondodendrine	Bis 49

Chondodendron microphyllum	d-Bebeerine	Bis 19
	Isochondodendrine	Bis 49
	L-Isococlaurine	(Benz.2)
Chondodendron platyphyllum	L-Berberine	Bis 18
	Isochondodendrine	Bis 49
	L-Isococlaurine	(Benz.2)
Chondodendron tomentosum	Cycleanine	Bis 48
	d-Chondocuranine	Bis 16
	d-Chondocurine	Bis 17
	Isochondodendrine	Bis 49
	d,l-Tubocurarine	Bis 25
	L-Curine	Bis 18
	Norcycleanine	Bis 50
Cissampelos insularis	Cycleanine	Bis 48
= Cyclea insularis	Cycleanoline	P13
= Paracyclea insularis	Insulanoline	Bis 51
	Insularine	Bis 52
	Isochondodendrine	Bis 49
	Magnoflorine	A 7
	Norcycleanine	Bis 50

Cissampelos pareira	Cissampareine	Bis 53
	Cyclanoline	P 13
	Isocondodendrine	Bis 49
	Hayatidine	Bis 21
	Hayatine	bis 22
	Hayatinine	Bis 23
	L-Curine	Bis 18
	4"-Omethylcurine	Bis 24
Cissampelos ochiaiana	Cyleanine	Bis 48
	Insularine	Bis 52
	Isochondodendrine	Bis 49
	L-Curine	Bis 18
Cissampelos mucronata	Isocondodendrine	Bis 49
Cocculus hirsutus	d,1-Coclaurine	Benz 1
	d-Trilobine	Bis 36
Cocculus laurifolius	Cocculidine	Misc.1
	Cocculine	Misc.2
	Dihydroerysodine	Misc.3
	Coclaurine	Benz.1
	Laurifoline	A 6

	Magnoflorine	A 7
	Trilobine	Bis 38
Cocculus leaeba	Palmatine	P 2
	Oxycanthine	Bis 71
Cocculus pendulus	1,2 -Dehydrokohatamine	Bis 58
(Guinaudeau, 1987)	1,2 -Dehydrokohatine	Bis 57
	1,2 -Dehydronortelobine	Bis 59
	5 -Hydroxyapateline	Bis 55
	5 -Hydroxytelobine	Bis 56
	Kohatamine	Bis 54
	Siddiquamine	Bis 61
	Siddiquiene	Bis 60
Cocculus sarmentosus	Cocsarmine	A1
	Isotriboline (homotriboline)	Bis 36
	Menisarine	Bis 42
	Trilobine	Bis 38
Cocculus trilobus	Cocculolidine	Misc.1
	Isotrilobine	Bis 36
	Magnoflorine	A7
	Normenisarine	Bis 43

	Trilobamine (daphnoline)	Bis 67
	Trilobine	Bis 32
Cyclea atjehensis	Cycleatjehenine	
	Cycleatjehine	
Cyclea barbata	Homoaromoline	Bis 68
	Isotetrandrine	Bis 8
Cyclea peltata	Berbamine	Bis 2
(Kupchan, 1973)	Cycleacurine	Bis 20
	Cycleadrine	Bis 3
	Cycleahomine chloride	Bis 4
	Cycleanorine	Bis 5
	Cycleapeltine	Bis 65
	Fangchinoline	Bis 6
	Isochondodendrine	Bis 49
Coscinium blumeanum	Berberine	P 1
	Jatrorrhizine	P 6
	Palmatine	P 2

Coscinium fenestratum	Berberine	P1
	Berberubine	P
	12,13-Dihydro-8-oxoberberine	P
	Jatrorrhizine	P 6
	N, N-Dimethyllindcarpine	A 10
	Oxyberberine	P
	Palmatine	P 2
	Tetrahydroberberine	P
	Thalifendine	P 15
Epinetrum cardifolium	Cycleanine	Bis 48
	Isochondodendrine	Bis 49
	Norcycleanine	Bis 50
Epinetrum mangenati	Cycleanine	Bis 48
	Isochondodendrine	Bis 49
	Norcycleanine	Bis 50
ibraurea chloroleuca	Berberine	P 1
(=F. tinctoria)	Berberrubine	P
(Siwon, 1981)	Dehydrocorydalmine	P 7
	Jatrorrhizine	P 6
	Magnoflorine	A 7
	Palmatrubine	P
	Pseudocolumbamine	P 20

Heptacyclum zenkeri	Dehydrodiscretine	P 17
Jatrorhiza palmata	Columbamine	P 4
	Jatrorrhizine	P 6
	Palmatine	P 2
Legnephora moorei	Dehydrocorydalmine	P 7
(=Pericampylus incanus)	Menisperine	2 1
	(N-methylisocorydinium)	
Limacia cuspidata	Cuspidaline	Bis 26
	Limacine	Bis 12
	Limacusine	Bis 69
Limacia oblongata	Cuspidaline	Bis 26
	Limacine	Bis 12
	Limacusine	Bis 69
Limaciopsis loangensis	Isotetrandrine	Bis 8
(Cave , 1978)	Nor-2 isotetrandrine	Bis 9
	N-Oxy-2 -isotetrandrine	Bis 10
	8-Oxopalmatine	P

Menispermum canadense	Dauricine Dehydrocheilanthifoline	Bis 27
Menispermum dauricum (Takani, 1983)	Acutumidine Acutumine Dauricine Daurinoline	- Bis 27 Bis 28
	Dauriporphine Menisporphine Sinomenine	A 17 A 18 M 1
Pachygone loyaltiensis	Apateline Bisnoraromoline Daphnandrine Daphnoline	Bis 34 - Bis 65 Bis 67
	1, 2 -Dehydroapateline 1, 2 -Dehydrotelobine Isotrilobine 0-Methylcocsoline	Bis 39 Bis 40 Bis 36 Bis 37

Pachygone ovata	Coclaurine	Benz.1
(Abdel-kawi, 1984)	Liriodenine	A 22
	Magnoflorine	A 7
	Nortrilobine	<u>_</u>
	Trilobine	Bis 38
Parabaena hirsuta	Palmatine	P 2
Pericampylus formosanus	(+) Stepharine	Pa3
Pleogyne australis	-Isochondodendrine	Bis 49
(=P. cuminghamii L.)	Curine	Bis 18
Pycnarrhena longifolia	Aromoline	Bis 62
	Berbacolorflammine	Bis 1
	Colorflammine	Bis 64
	Daphnoline	Bis 67
	Homoaromoline	Bis 68
	Krukovine	Bis 11
	Limacine	Bis 12
	Magnoflorine	A 7
	Obaberine	Bis 13
	Pycnarrhine	Misc. 7

Table 1 (continue)

Pycnarrhena manillensis	Berbamine	Bis 2
	Isotetrandrine	Bis 8
	Phaeanthine	bis 14
	Pycnamine	Bis 15
Pycnarrhena novoguinensis	Berbamine	Bis 2
(Verpoorte, 1978)	Isofangchinoline	Bis 7
	Limacine	Bis 12
	Phaeantine	Bis 14
	Pycnamine	Bis 15
Sinomenium acutum	Disinomenine	M 2
	Magnoflorine	A 7
	Michelalbine	A 9
	Norsinoacutine	м 3
	Sinactine	P 11
	Sinoacutine	M 4
	Sinomenine	M 1
	Stepharine	Pa 3
	Tuduranine	A 16

Table 1 (continue)

Stephania capitata	Crebanine	A 3		
	Cycleanine	Bis 48		
	d-Dicentrine	A 4		
	Epistephanine	Bis 72		
	Phanostenine	A 11		
	Stephanine	A 14		
Stephania cepharantha	Berbamine	Bis 2		
	Cepharamine	н 1		
	Cepharanthine	Bis 63		
	Cycleanine	Bis 48		
	Homoaromoline	Bis 68		
	Isotetrandrine	Bis 8		
Stephania drinklagei	(+) Corydine	A 2		
	Dicentrine	A 4		
	(+) Isocorydine	A 5		
	(-) Roemerine	A 12		
Stephania elegans	Aknadinine	M 5		
	Cyclanoline	P 13		
	Cycleanine	Bis 48		
	Epihernandolinol	м 6		
	Hasubanonine	н 4		

	Isochondodendrine	Bis 49
	Isotetrandrine	Bis 8
	Magnoflorine	A 7
	N-Methylcorydalmine	P 16
		P 4
Stephania glabra	Columbamine	
	(-) Corydalmine	P 8
	Cycleanine	Bis 48
	Dehydrocorydalmine	P 7
	Gindarine = Tetrahydropalmatine	P 3
	Gindarinine = Palmatine	P 2
	Jatrorrhizine	P 6
	Palmatine	P 2
	Stepharanine	P 9
	Stepharatine	P 14
	(-) Stepholidine	P 10
	Tetrahydropalmatine	P 3
Stephania hernandifolia	4-Demethylhasubanonine	Н 2
	4-Demethylnorhasubanonine	Н 3
	Isochondodendrine	Bis 49
	Fangchinoline	Bis 6
	Isotrilobine	Bis 36

tephania japonica	Cyclenoline	P 10
	Cyclanoline	P 13
(Matsui, 1978,1982,1984.	Epistephamiersine	H 12
Yamamura, 1985)	Epistephanine	-
	Hasubanonine	H 4
	Homostephanoline	Н 5
	Hypoepistephanine	Bis 73
	Insularine	Bis 52
	Lanuginosine	A 23
	Magnoflorine	A 7
	Metaphanine	Н 6
	Miersine	Н 18
	Oxo-epistephamiersine	H 14
	16-Oxoprometaphanine	н 8
	Oxostephabinine	Н 10
	Oxostephamiersine	Н 13
	Oxostephanine	A 10.
	Oxostephasunoline	Н 15
	Protostephanine	Misc 4
	Prometaphanine	н 7
	Stebisimine	Bis 74
	Stephamiersine	Н 11
	Stephanine	A 14
	Stephasunoline	H 15

	Stepinonine	Misc 5		
	Steponine	P 12		
	Thalrugosine	-		
Stephania kwansiensis	Tetrahydropalmatine	P 3		
Stephania longa	Longanone	Н 17		
Stephania sasakii	Berbamine	Bis 2		
(Kumitomo, 1981)	Bisakanadinine	M 7		
	Cepharanthine	Bis 63		
	Crebanine	A 3		
	Dehydrocrebanine	A 19		
	Dehydrostesakine	A 20		
	4,5 - Dioxydehydrocrebanine	A 21		
	4-Hydroxycrebanine	A 31		
	d-Isocorydine	A 5		
	Lanuginosine	A 23		
	Liorodenine	A 22		
	L-Tetrahydropalmatine	P 3		
	N-Methylpapaveralinium	Benz.6		
	Phanostenine	A 11		
	(R) -Roemeroline	A 12		

	Steponine	P 12		
	Stesakine	A 15		
Stephania suberosa	Cepharanthrine 2'N-oxide			
(Patra, 1986)	2-norcepharanthine			
	Norstephasubine	Bis 76		
	Stephasubinine	Bis 77		
	Stephasubine	Bis 75		
Stephania tetrandra	Cyclanoline	P 13		
	Fanchinoline	Bis 6		
Stephania venosa	Anonaine	A		
	Apoglazionine	A		
	Asimilobine	A		
	Ayuthianine	A 28		
	Corydine	A 2		
	Crebanine	A 3		
	Kikumamanin	P		
	Mecambroline	A		
	N-Carboxamidostepharine	Pa 1		
	Nuciferline	A		
	O-Methylstepharinosine	Pa 4		
	Oxostephanosine	A 25		

		T
	Reticuline	Benz.5
	StephadiolamineN-oxide	<u>-</u>
	Stepharine	Pa3
	Stepharinosine	Pa5
	Sukhodianine	A 29
	Tetrahydropalmatine	P 3
ta garage	Thailandine	A 26
	Thalrugosamine	-
	Tuduranine	A 16
	Ushinsunine	A 30
	Uthongine	A 27
ynclisia scabrida	Cocsuline	Bis 35
(Ohiri, 1983)	Cycleanine	Bis 48
	Norcycleanine	Bis 50
'iliacora acuminata	Tiliacorine	Bis 31
(=T. racemosa)	Tiliarine	Bis 30
	Tiliacorenine	Bis 32
iliacora dinklagei	Nortiliacorinine A	Bis 33
	Tiliacorine	Bis 31
	Tiliacorinine	Bis 32

Tiliacora triandra	Nortiliacorinine A	Bis 33		
	Tiliacorine	Bis 31		
	Tiliacorinine	Bis 32		
	Tilianangine	Bis 47		
	Yanangine	Bis 46		
Tinomiscium petiolare	L-Isocorypalmine	P 5		
Tinospora bakis ^	Berberine /	P 1		
(=Coculus bakis)	Palmatine	P 2		
Tinospora capillipes	Columbamine	P 4		
(Chang, 1984)	Dehydrodiscretamine	P 17		
	Jatrorrhizine	P 6		
	Palmatine	P 2		
	Stepharamine	P 9		
Tinospora crispa/	Berberine	P 1		
(=Cocculus crispa)	Palmatine	P 2		
Tinospora rumphii	Berberine/	P 1		

Table 1 (continue)

Trilisia dictyophyllia	Cocsuline	Bis 35
(Spiff, 1981)	Tridictophylline	M 8
	Trigilletimine	Bis 41
Trilisia gilletti	Gilletine	Bis 44
	Isogelletine-N-oxide	Bis 45
	Obamegine	Bis 13
	Stebisimine	Bis 74

Note: P = Protoberberine alkaloids, which can find their structures in table 3 (page 71-113).

Table 2 The structure of formulae in table 1

Aporphine a	B 6a 6a 7	75 N CH o					
Alkaloid	1	2	6	8	9	10	11
Cocsarmine (A1)	OMe	ОМе	(Me) ₂	-	OMe	OMe	-
Corydine (A2)	ОН	OMe	Me	-	-	OMe	OMe
Crebanine (A3)	-o-c	-0-CH ₂ -0-		OMe	OMe	-	-
Dicentrine (A4)	-0-0	H ₂ -0-	Me	-	OMe	OMe	-
Isocorydine (A5)	OMe	OMe	Me	-	-	OMe	ОН
Laurifoline (A6)	ОН	OMe	(Me) ₂	-	ОН	OMe	-
Magnoflorine (A7)	ОН	OMe	(Me) ₂	-	-	OMe	ОН
Menisperine (A8)	OMe	OMe	(Me) ₂	-	-	OMe	ОН
Michelalbine (A9)	-0-C	-0-CH ₂ -0-		-	-	-	-
N, N-dimethyllindcarpine (A10)	OMe	ОН	(Me) ₂	-	-	OMe	ОН
Phanostenine (A11)	-о-с	-0-CH ₂ -0-		-	OMe	ОН	-
Roemerine (A12)	-o-c	-0-СН2-0-		-	-	-	-
Roemeroline (A13)	-o-c	-o-cH ₂ -o-		-	ОН	-	
Stephanine (A14)	-o-c	-о-сн ₂ -о-		OMe	-	-	-
Stesakine (A15)	-0-C	H ₂ -0-	Me	OMe	ОН	- 0	-
Tuduranine (A16)	OMe	ОМе	Н	-	-	ОН	-

Dauriporphine (A17) R=OMe

Menisporphine (A18) R=H

Dehydrocrebanine (A19) R=Me

Dehydrostesakine (A20) R=H

4,5-Dioxydehydrocrebanine (A21)

Lioriodenine (A22) R_1 =H R_2 =H Lanuginosine (A23) R_1 =H R_2 =OMe Oxostephanine (A24) R_1 =OMe R_2 =H Oxostaphanosine (A25) R_1 =OH R_2 =H Thailandine (A26) R_1 =OMe R_2 =H N-Me Uthongine (A27) R_1 =OMe R_2 =OMe N-Me

$$\bigcap_{0} \bigcap_{\mathbb{R}_{2}} \mathbb{R}_{1}$$

Ayuthianine (A28) R_1 =OMe R_2 =H Sukhodianine (A29) R_1 =OMe R_2 =OMe Ushinsunine (A30) R_1 =H R_2 =H

4-Hydroxycrebanine (A31)

1-Benzyl-tetrahydroisoquinoline (Benz)

Alkaloid	R	3'	4'	6	7
Coclaurine (Benz 1)	н	-	ОН	OMe	ОН
Isococlaurine (Benz 2)	н	-	ОН	ОН	OMe
Laudanine (Benz 3)	Me	ОН	OMe	OMe	OMe
N-methylcoclaurine (Benz 4)	Me	-	ОН	OMe	ОН
Reticuline (Benz 5)	Ме	ОН	OMe	ОМе	ОН

N-Methylpapaveraldinium chloride

(Benz 6)

Table 2 (continue)

Bisbenzylisoquinoline alkaloids (Berbamine type)

Alkaloids	R ₂	R ₃	R ₄	R ₅	R'2	R'3	Config	uration
Berbacolorflammine (Bis 1)	Me	Ме	Н	Me	Ме	Ме	-	
Berbamine (Bis 2)	Ме	Ме	Ме	Н	Me	Ме	1-R,	1'-S
Cycleadrine (Bis 3)	Ме	Ме	Н	Ме	Me	Me	-	-
Cycleahomine Cl (Bis 4)	(Me) ₂	Ме	Ме	Me	Ме	Me	1-S,	1'-8
Cycleanorine (Bis 5)	Me	Ме	Ме	Me	Н	Me	1-S,	1'-S
Fanchinoline (Bis 6)	Me	Me	Н	Ме	Ме	Ме	1-S,	1'-S
Isofanchinoline (Bis 7)	Me	Me	Н	Ме	Me	Me	1-S,	1'-S
Isotetrandrine (Bis 8)	Me	Me	Me	Ме	Me	Me	1-R,	1'-S
Nor-2-Isotetrandrine (Bis 9)	Me	Ме	Me	Me	Н	Me	1-R,	1'-S
N-Oxy-2'-isotetrandrine								
(Bis 10)	Ме	Ме	Me	Me	O,Me	Me	1-R,	1'-8
Krukovine (Bis 11)	Ме	Me	Н	Н	Me	Me	1-R,	1'-R

Table 2 (continue)

							1, 18, 1	
Limacine (Bis 12)	Ме	Me	Н	Ме	Me	Me	1-R,	1'-R
Obamegine (Bis 13)	Ме	Ме	Н	Н	Ме	Ме	1-R',	1'-8
Phaeanthine (Bis 14)	Ме	Ме	Me	Ме	Ме	Ме	1-R,	1'-R
Pycnamine (Bis 15)	Me	Ме	Ме	Н	Me	Me	1-R,	1'-R

Bisbenzylisoquinoline alkaloids (Curine type)

Alkaloid	R ₂	R ₃	R ₄	R ₅	R'2	R'3	Config	uration
d-Chondocurarine (Bis 16)	(Me) ₂	Ме	Н	Н	(Me) ₂	Me	-	-
d-Chondocurine (Bis 17)	Ме	Me	Н	Н	Ме	Ме	-	-
1 -Curine (Bis 18)	Me	Me	Н	Н	Ме	Me	1-S,	1'-S
d-Curine (Bis 19)	Me	Me	Н	Н	Me	Ме	1-R,	1'-R
Cycleacurine (Bis 20)	Me	Me	Н	Н	Ме	Н	1-R,	1'-R
Hayatidine (Bis 21)	Me	Ме	Н	Ме	Me	Me	1-S,	1'-R
Hayatine (Bis 22)	Ме	Me	Н	Н	Me	Me	_	-
Hayatinine (Bis 23)	Me	Me	Н	Me	Me	Me	_	_

4"-0-Methylcurine (Bis 24)	Me	Me	Н	Me	Me	Me	1-S,	1'-S
d-Tubocurarine (Bis 25)	(Me) ₂	Ме	Н	Н	(Me) ₂	Me	_	-

Note: Curine = Bebeerine = Chondodendrine

Bisbenzylisoquinoline alkaloids (Dauricine type)

Alkaloid	R ₂	R ₃	R ₄	R ₅	R'2	R'3	R'4	Config	uration
Cuspidaline (Bis 26)	Me	Ме	Н	Me	Ме	Me	Н	1-R,	1'-R
Dauricine (Bis 27)	Ме	Me	Ме	Н	Ме	Me	Ме	1-R,	1'-R
Daurinoline (Bis 28)	Ме	Ме	Ме	Н	Me	н	Me	1-R,	1'-R
Lindoldhamine (Bis 29)	Н	Ме	Н	Н	Н	Me	Н	1-R,	1'-R

Table 2 (continue)

Bisbenzylisoquinoline alkaloids (Dibenzo-p-dioxin type)

Alkaloid	R ₂	R ₃	R ₄	R'2	R'3	Config	uration
Tiliarine (Bis 30)	Ме	Me	Н	Н	Me	1-S,	1'-S
Tiliacorine (Bis 31)	Me	Me	Me	Ме	Н	1-R,	1'-8
Tiliacorinine (Bis 32)	Me	Me	Me	Ме	Н	1-S,	1'-8
Nortiliacorinine A (Bis 33)	Н	Ме	Me	Ме	Н	1-S,	1'-S

$$\begin{array}{c|c} R_2 & R_3 & \\ \hline \\ R_1 & \\ \hline \\ OR_3 & \\ \hline \end{array}$$

Alkaloid	R ₂	R ₃	R'2	R'3	Config	guration
Apateline (Bis 34)	Н	Н	Me	Me	1-R,	1'-S
Cocsuline (Bis 35)	Me	Н	Me	Me	1-S,	1'-S
Isotrilobine (Bis 36)	Ме	Me	Ме	Me	1-S,	1'-S
O-Methyl cocsoline (Bis 37)	Н	Ме	Me	Me	1-S,	1'-S
Trilobine (Bis 38)	Ме	Me	н	Ме	1-S,	1'-S

1,2-Dehydroapateline (Bis 39)

1,2 -Dehydrotelobine (Bis 40)

Trigilletimine (Bis 41)

$$H_3C$$

$$OCH_3$$

$$OCH_3$$

$$OCH_3$$

Menisarine (Bis 42)

Normenisarine (Bis 43)

(=0-demethylmenisarine, exact location of OH is not determind)

Gilletine (Bis 44)

Isogilletine-N-Oxide (Bis 45)

Yanangine (Bis 46)

Tilianangine (Bis 47)

Table 2 (continue)

Bisbenzylisoquinoline alkaloids (Isochondodendrine type)

$$R_2$$
 R_1
 OR_3
 OR_4
 OR

Alkaloid	R ₂	R ₃	R ₄	R'2	R'3	R'4	Config	uration
Cycleanine (Bis 48)	Me	Me	Me	Me	Me	Ме	1-R,	1'-R
Isochondodendrine (Bis 49)	Ме	Ме	Н	Ме	Ме	Н	1-R,	1'-R
Norcycleanine (Bis 50)	Ме	Ме	Ме	Ме	Ме	Н	-	-

$$R_2$$
 N
 OR_4
 OR_4

Alkaloid	R ₂	R ₃	R ₄	R'2	R'3	Configuration
Insulanoline (Bis 51)	Ме	Me	Н	Me	Me	1-R, 1'-R
Insularine (Bis 52)	Ме	Ме	Ме	Ме	Me	1-R, 1'-R

$$\begin{array}{c|c} & & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & &$$

Cissampareine (Bis 53)

Table 2 (continue)

Bisbenzylisoquinoline alkaloids (Micranthine type)

$$\mathbb{R}_2 = \mathbb{R}_1$$

$$\mathbb{R}_2$$

$$\mathbb{R}_1$$

$$\mathbb{R}_2$$

$$\mathbb{R}_1$$

$$\mathbb{R}_2$$

Alkaloid	R ₂	R ₃	R'2	R'3	Configuration
Kohatamine (Bis 54)	Н	Ме	Ме	Me	1-S, 1'-S
5'-Hydroxyapateline (Bis 55)	Н	Н	Me	Me	1-R, 1'-S
5'-Hydroxytelobine (Bis 56)	Н	Me	Me	Me	1-R, 1'-S

Alkaloid	R ₁	R'2	R'3	R'4	Con	figuration
1,2-Dehydrokohatine (Bis 57)	Н	Ме	Ме	ОН	_	1'-S
1,2-Dehydrokohatamine (Bis58)	Ме	Ме	Me	ОН	-	1'-S
1,2-Dehydro-2'-nortelobine (Bis 59)	Ме	Н	Ме	Н	-	1'-S
	Simon					

Alkaloid	R	R'2	R'3	R'4	Configuration
Siddiquine (Bis 60)	Н	Ме	Ме	ОН	- 1'-S
Siddiquamine (Bis 61)	Ме	Me	Ме	ОН	- 1'-S

Table 2 (continue)

Bisbenzylisoquinoline alkaloids (Oxyacanthin type)

$$R_2 \xrightarrow[R_1]{\text{OR}_3} R_3^{\prime} O$$

$$R_4^{\prime} O$$

$$OR_4 O$$

$$OR_4 O$$

Alkaloid	R ₂	R ₃	R ₄	R'2	R'3	R'4	Configuration
A							
Aromoline (Bis 62)	Me	Ме	Н	Ме	Ме	Н	1-R, 1'-S
Cepharanthine (Bis 63)	Ме	Ме	Me	Ме	- CH	H ₂ -	1-R, 1'-S
Colorflamming (Bis 64)	Me	Me	Ме	Ме	Ме	Н	
Cycleapeltine (Bis 65)	Me	Ме	Me	Ме	Me	Н	1-S, 1'-S
Dapnandrine (Bis 66)	Н	Me	Ме	Ме	Ме	Н	1-R, 1'-R
Dapnoline (Bis 67)	Н	Ме	Н	Me	Me	Н	1-R, 1'-S
Homoaromoline (Bis 68)	Me	Ме	Me	Ме	Me	Н	1-R, 1'-S
Limacusine (bis 69)	Me	Me	Me	Ме	Me	Н	1-R, 1'-R
Obaberine (Bis 70)	Me	Me.	Me	Ме	Me	Ме	1-R, 1'-S
Oxyacanthine (Bis 71)	Ме	Ме	Н	Ме	Me	Me	1-R, 1'-S

$$H_3C$$
 H_3CO
 H_3CO
 OCH_3
 H_3CO
 OCH_3
 H_3CO
 OCH_3
 OCH_3

R=Me Epistephanine (Bis 72)

R=H Hypoepistephanine (Bis73)

Stebisimine (Bis 74)

R=Me, Stephasubine (Bis 75)

R=H, 2-Norstephasubine (Bis 76)

Stephasubinine (Bis 77)

Hasubanan alkaloids (H)

Alkaloid	3	4	8	R
Cepharamine (H1)	OMe	ОН	Н	Me
4-Demethylhasubanonine (H2)	OMe	ОН	OMe	Ме
4-Demethylnorhasubanoline (H3)	OMe	ОН	0Me	Н
Hasubanonine (H4)	OMe	OMe	OMe	Me
Homostephanoline (H5)	ОН	OMe	OMe	Ме

1 8	H ₃ CO~			
	H ₃ CO			
	3	16		
		O N-CH	13	
		OH 7		

Alkaloids	7	16
Metaphanine (H6)	0	Н2
Prometaphanine (H7)	OMe	Н2
16-Oxoprometaphanine (H8)	ОМе	0

Stephabenine (H9) $R = H_2$

Oxostephabenine (H10) R = O

H ₃ CO		
H ₃ CO		
N-CH ₃		
OCH3		
7 0013		
Alkaloids	7	16
tenhamiergine (H11)	OMe	H ₂
tephamiersine (H11) pistephamiersine (H12)	OMe	Н2
		H ₂

Alkaloids	3	4	6	7	8	16
Stephasunoline (H15)	OMe	ОМе	ОН	0Me	ОН	Н2
Longanone (H16)	OMe	ОН	0	0Me	ОМе	Н2
Oxostephasunoline (H17)	OMe	ОМе	ОН	OMe	ОН	0
Miersine (H18)	OMe	ОМе	ОН	OMe	ОН	H ₂

Proaporphine alkaloids (Pa)

Alkaloid	1	2	6(R)
N-Carboxamidostepharine (Pal)	OMe	ОМе	-conh ₂
Pronuciferine (Pa2)	OMe	0Me	Ме
Stepharine (Pa3)	OMe	OMe	Н

O-methylstapharinosine (Pa4)

Stepharinosine (Pa5)

Morphinondienone alkaloids (M)

$$_{\text{OCH}_3}^{\text{H}_3\text{CO}}$$

Sinomenine (M1)

(Coculine)

$$H_3$$
CO
 OCH_3
 OCH_3

Disinomenine (M2)

(Dehydrosinomenine)

Norsinoacutine R=H (M3)

Sinoacutine R=Me (M4)

$$H_3CO$$
 RO
 OCH_3
 OCH_3

Aknadinine (M5)

Miscellaneous base (Misc.)

Cocculidine (Misc 1) R_1 =OMe, R_2 =OMe

Cocculine (Misc 2) R_1 =OH , R_2 =OH

Dihydrocrysodine (Misc 3) R_1 =OMe, R_2 =OH

Protostephanine (Misc 4)

$$H_3C$$
 OCH_3
 OCH_3

Stepinonine (Misc 5)

Pycnarrhina (Misc 7)

$$_{\mathrm{Ho}}$$
 $_{\mathrm{OCH_{3}}}$ $_{\mathrm{OCH_{3}}}$

Bisnorargemonine (Misc 6)

2.2 Distribution of protoberberine alkaloids in plants

The protoberberines are one of the most widely distributed of the isoquinoline alkaloid groups, being present in at least nine plant families particularly the Annonaceae, Berberidaceae, Lauraceae, Menispermaceae, Papaveraceae and Rutaceae. Most protoberberine alkaloids exist in nature either as tetrahydroprotoberberines or quaternary protoberberine salts, but some dihydroprotoberberines are also known.

The basic structures of these three subgroups are mentioned below.

Dihydroprotoberberines

Tetrahydroprotoberberines Quaternary protoberberine salt

Substituents are usually present at C-2 and C-3 and either at C-9 and C-10 or at C-10 and C-11. In the case of C-10 and C-11 are substituted it is called "pseudo" variety. In some instances a hydroxyl or methoxyl substituent may present at C-1. A methyl group is sometimes found at C-8 or C-13, while in a few cases an alcoholic hydroxyl is located at C-13 or C-5. Some retroprotoberberines are known which are characterized by the presence of one extra carbon atom as a side chain bonded to ring D. (Shamma, 1972)

The distribution of protoberberine alkaloids in plants are summarized as table 3.

Table 3 Distribution of protoberberberine alkaloids in plants

3.1 0	uater 3 2	nary	proto		erine 9 10
Name	2	Strue	cture	10	Plants
Berberine (Umbellatine)	-0-CF	H ₂ -0-	OMe	OMe	Anamirta cocculus Aquilegia olympica Arcangelisia flava A. lemniscata A. lourerie Argemone mexicana Berberis asiatica B. buxifolia B. darwinii B. empetrifolia B. glauca B. julianae

	ГТ	T
Berberine		B. lycium
		B. oblonga
		B. nervosa
		B. petiolaris
		B. vulgaris
		Chelidonium majus
		Conting drawnlanding
		Coptis groenlandica C. japonica
		C. occidentalis
		C. quinavefolia
		C. teeta
		C. trifolia
		Corydalis cheilantheifolia
*		C. ophicarpa
		Coscinium blumeanum C. fenestratum
. // / / / / / / / / / / / / / / / / /		C. wallichianum
		Dicranostigma lactucoides
		D. leptopodum
у Э		Evodia meliifolia
		Fibraurea chloroleuca
		F. tinctoria

		Fumaria kraliki
		Glaucium corniculatum
12 12 1		
2 21		G. grandiflorum
A P I		Hunnemannia fumariaefolia
		Hydrastis canadensis
		nyarastis canadensis
		Mahonia aquifolium
		M. repens
		Nandina domestica
		Papaver albiflorum
		P. lecoquii
		P. rhoeas
		Phellodendron amurense
		P. wilsonii
		The liet was a laisure
		Thalictrum alpinum
		T. baicalense
		T. foliosum
		m iamai
		T. javanicum
		T. longistylum
		T. lucidum
		T. minus
		1. minus
		T. polyganum
		T. revolutum
1 - 1		Tinognore hearzideni
		Tinospora baenzigeri
		T. crispa
		T. glabra
	1	

Table 3 (continue)

					T. merrilliana T. rhumphii T. smilacina Toddalia aculeata Xylopia polycarpa Zanthoxylum caribaeum Z. monophyllum
Berberubine	-0-C	H ₂ -0-	ОН	OMe	Coscinium fenestratum Fibraurea chloroleuca Thalictrum polygranum
Columbamine	ОН	ОМе	OMe	OMe	Anamirta cocculus Arcangelisia flava Berberis oblonga B. vulgaris Burasaia madagascariensis Chasmanthera dependens Coptis quinavefolia Enantia chlorantha E. polycarpa Fibraurea chloroleuca Jatrorrhiza palmata Mahonia repens Stephania glabra

Table 3 (continue)

Γ			r
			Thalictrum alpinum T. folialosum T. japonicum T. longistylum T. minus T. polycarpum T. revolutum T. rugosum
Coptisine	-O-CH ₂ -O-	-O-CH ₂ -O-	Chelidonium majus Coptis groenlandica C. japonica C. quinavefolia Corydalis cava C. ophiocarpa Dicranostigma lactucoides D. leptopodum Fumaria densiflora F. judaica
			F. kraliki F. parviflora Hunnemannia fumariaefolia Papaver albiflorum P. conifine P. lecoquii

Table 3 (continue)

					P. rupifragum P. pseudo-orientale
Dehydro- cheilanthifoline	ОН	OMe	-0-СН2-0-		Bocconia cordata Corydalis ophiocarpa Fumari parviflora Menispermum cannadense
Dehydrocorydalmine	OMe	OMe	OMe	ОН	Arcangelisia flava Berberis floribunda Corydalis ambigna Legnephora moorii Stephania glabra
Dehydrodiscretamine	OMe	ОН	ОМе	ОН	Corydalis tashiroi Thalictrum foliolosum
Demethylleneberberine	ОН	OH	OMe	OMe	Thalictrum javanicum
Groenlandicine	OMe	ОН	-0-0	CH ₂ -O-	Bocconia cordata Coptis groenlandica Coptis spp. Nandina domestica

Table 3 (continue)

atrorrhizine	OMe	ОН	OMe	OMe	Arcangelisia flava
(Jateorhizine,					A. lourerii
Neprotine)					Berberis julianae
					B. heteropoda
					B. oblonga
					B. vulgaris
					Burasaia madagascariensis
					Chasmanthera dependens
					Coptis japonica
					C. quinavefolia
					Coscinium blumeanum
					C. fenestratum
					C. wallichianum
					Enantia chlorantha
					E. polycarpa
					Fibraurea chloroleuca
					Jatrorrhiza palmata
					Mahonia philippinensis
					M. repens
					Stephania glabra
					Thalictrum foliolosum
					T. lucidum
					T. revolutum
					Tinospora baenzigeri
					T. crispa

Table 3 (continue)

Palmatine	OMe	OMe	OMe	OMe	Anamirta cocculus
				111	Arcangelisis flava
					A. lourerii
					Berberis jaeschkeana
					B. julianae
					B. oblonga
					B. petiolaris
					B. vulgaris
					Chasmanthera dependens
					Cocculus carolinus
					C. leaeba
					Coscinium blumeanum
					C. fenestratum
					C. wallichianum
					Enantia chlorantha
					E. pilosa
					E. polycarpa
					Fibraurea chloroleuca
					Fumaria densiflora
					Jatrorrhiza palmata
					Leontice leontopatalum
					Mahonia repens
					Papaver pseudo-orientale
					Stephania glabra
					S. kwansiensis

Table 3 (continue)

					Tinospora cordifolia
					T. crispa
					T. glabra
					T. merrilliana
					T. sagittata
					T. sinensis
					Thalictrum alpinum T. foliolosum
					T. javanicum
					T. longistylum
		Commission of the Commission o			T. minus
					T. podocarpum
					T. revolutum
Palmatrubine	OMe	0Me	ОН	OMe	Fibraurea chloroleuca
8 4 1					Stephania glabra
					Thalictrum polygamum
Stepharanine	ОН	OMe	OMe	ОН	Stephania glabra
					S. intermedia
			-		Tinospora capillipes
Thalifendine	-0-C	H ₂ -0-	OMe	ОН	Arcangelisia flava
					Thalictrum alpinum
					T. fendleri
		-	1		

		T. foliolosum
		T. minus
		T. podocarpum
		T. polygamum
		T. revolutum
		T. rugosum

3.2 Quaternary pseudoprotoberberines

Name		Stru	cture		Plants
	2	3	10	11	
Dehydrodiscretine (Pseudojatrorrhizine)	ОМе	ОН	ОМе	ОМе	Fibraurea chloroleuca Heptacyclum zenkeri Thalictrum fauriei

Dehydropseudo cheilanthifoline	ОН	OMe	2		Corydalis ophiocarpa Fumaria parviflora
one i an en i o i ne					Isopyrum thalictroides
Pseudoberberine	-O-C	H ₂ -0-	OMe	OMe	Isopyrum thalictroides
Pseudocolumbamine	ОН	ОМе	OMe	ОМе	Isopyrum Thalictroides Fibraurea chloroleuca
Pseudocoptisine	-0-C	н ₂ -0-	-0-C	Н2-0-	Coptis groelandica Isopyrum thalictroides
Pseudopalmatine	OMe	ОМе	ОМе	OMe	Enantia polycarpa
Thalifaurine	OMe	ОН	-0-C	H ₂ -0-	Thalictrum fauriei

3.3 Quaternary protoberberines methylated at C-13 $^{\circ}$

Table 3 (continue)

Name		St	ructu	re		
иаме	2	3	9	10	13	Plants
Corysamine	-o-c	Н2-0-	-0-с	н ₂ -0-	Ме	Chelidonium majus Corydalis incisa C. cava C. lutea Dicranostigma lactucoides D. leptopodum Hunnemannia fumariaefolia Papaver albiflorum P. rupifragum
Dehydroapocavidine	ОН	ОМе	-0-C	н ₂ -о-	Me	Corydalis cava
Dehydrocavidine	ОМе	ОМе	-0-C	н ₂ -0-	Ме	Corydalis cava Corydalis meifolia
Dehydrocorydaline	ОМе	ОМе	OMe	ОМе	Ме	Corydalis ambigua C. decumbens C. tuberosa Berberis floribunda

3.4 Quaternary pseudoprotoberberines methylated at C-13

Name		. St	ructu	re		Plants
	2	3	10	11	13	
Worenine	-0-C	H ₂ -0-	-0-CI	H ₂ -0-	Ме	Coptis chinensis

3.5 Quaternary protoberberine hydroxylated at C-5

Name		Struct	ure		
Nume	2 3	5	9	10	Plants
Berberastine	-O-CH ₂ -	О- ОН	OMe	OMe	Coptis spp. Hydrastis canadensis
Thalidastine	-0-СН2-	0- ОН	OMe	OH	Thalictrum fendleri T. foliolosum T. minus T. rugosum

3.6 Tetrahydroprotoberberines

Name		Stru	cture		Plants
	2	3	9	10	riants
(-) Aequaline	OMe	ОН	OMe	ОН	Desmos tiebaghiensis
((-) Discretamine)					Duguetia calycina
					Mitrella kentii
					Schefferomitra subaequalis Xylopia buxifolia
					X. discreta
Bharatamine'	ОН	ОМе	-	-	Alangium lamarckii
Canadine	-0-C	H ₂ -0-	ОМе	OMe	Corydalis cava C. cheilantheifoline

Table 3 (continue)

	,			
				Corydalis ophiocarpa C. ternata C. tuberosa Fagara rhoifolia Hydrastis canadensis Mahonia aquifolium Papaver albiflorum P. lecoquii Xanthoxylum brachyacanthum X. veneficum
			7	A. Venericam
Cheilanthifoline	ОН	OMe	-0-CH ₂ -0-	Argemone spp. Cheilidonium meifolia Corydalis cheilanthifolia C. ophiocarpa C. scouteri C. sibirica Fumaria judaica F. officinalis F. parviflora
				F. vaillantii Papaver macrostomum P. trinifolium

Table 3 (continue)

Corydalmine	OMe	OMe	OMe	ОН	Corydalis pallida
(Kikemanine,					Papaver oligosperma
Schefferine,					Polyalthia oligosperma
Cycemanine)		The second second			Schefferomitra subaequalis
					Stephania glabra
					S. Suberosa
Corypalmine	OMe	ОН	OMe	OMe	Berberis floribunda
(discretinine,					B. hemaloica
Tetrahydroja-					Coptis teeta
trorrhizine)					Corydalis caseana
					C. cava
					C. cheliantheifolia
					C. incisa
					C. ochroleuca
					C. ophiocarpa
					C. thalictrifolia
					C. tuberosa
					Dicentra oregana
					Guatteria discolor
					Hydrastis canadensis
					Mahonia aquifolium
					Pachypodanthium confine
					P. staudtii
					Xylopia discreta

Table 3 (continue)

Isocorypalmine	ОН	OMe	OMe	OMe	Bocconia frutescens
(Tetrahydro-					Corydalis cava
columbamine)					
, , , , , , , , , , , , , , , , , , , ,					C. lutea
					C. ophiocarpa
					Glaucium fimbrilligerum
					Hydrastis canadensis
					Pachypodanthium confine
					P. staudtii
					Tinomiscium petiolare
Nandinine	-0-0	H ₂ -0-	ОН	OMe	N
		412 ⁻⁰⁻	OH	Оме	Nandina domestica
Sinactin	OMe	ОМе		п о	
	OMe	Оме	-0-0	н2-0-	Corydalis meifolia
					Fumaria officinalis
					Sinomenium acutum
Scoulerine	ОН	OMe	ОН	0Me	Bocconia frutescens
					Corydalis cava
					C. caseana
					C. micrantha
					C. montana
					C. sibirica
					C. tuberosa
					Cryptocarya longifolia
					JPoolar Ja Toligi10118

Table 3 (continue)

			Erythrina orientalis Eshcholtzia lobbii Fumaria officinalis Hunnemannia fumariaefolia Hypecoum procumbens Papaver albiflorum
			P. lecoquii P. tauricola P. trinifolium
Stepholidine	OH OMe	OMe OH	Desmos tiebaghiensis Monanthotaxis cauliflora Stephania glabra S. suberosa
Stylopine (Tetrahydrocoptisine)	_	-0-CH ₂ -0-	Cheilidonium gortschakovii C. majus C. meifolia Corydalis cava C. marschallia C. ophiocarpa C. solida Fumaria judaica F. kraliki

Table 3 (continue)

					Fumaria parviflora F. schleicheri F. vaillantii Papaver albiflorum P. lecoquii P. rupifragum
Tetrahydropalmatine	OMe	OMe	OMe	OMe	Chasmanthera dependens
(Gindarine,					Corydalis ambigua
Caseanine)					C. aurea
					C. caseana
					C. cava
					C. decumbens
					C. lutea
					C. micrantha
					C. montana
					C. nobilis
					C. ochraleuca
					C. pallida
					C. platycarpa
					C. tuberosa
					Fibraurea chloroleuca
					Glaucium vitellinum
					Hydrastis canadensis

Table 3 (continue)

					Leontica leontopatalum Pachypodanthium confine Stephania glabra S. kwansiensis S. sasakii S. suberosa
Tetrahydropalmatrubine	OMe	0Me	ОН	OMe	Stephania suberosa
Tetrahydrothalifendine	-0-C	н ₂ -о-	ОМе	ОН	Thalictrum fendleri

${\tt 3.7\ Tetrahydropseudoprotoberberines}$

Name		Stru	cture		Plants
	2	3	9	10	
Artavenustine	OMe	ОН	ОН	ОН	Annona montana A. muricata
Coreximine (Coramine)	ОН	OMe	OMe	ОН	Artabotrys venustus Asimina triloba Chasnanthera dependens Corydalis spp. Dicentra eximia Erythrina orientalis Guatteria ouregou Papaver somniferum Stephania suberosa

Table 3 (continue)

Coryovanine (Corygoranine)	ОН	OMe	-0-0	H ₂ -0-	Corydalis govaniana Stephania suberosa
(()				T	
Corytenchine	OMe	ОМе	OMe	ОН	Corydalis ochotensis
10-Demethylxylopinine	OMe	0Me	ОН	OMe	Duguetia calycina
10-Demethyldiscretine	ОМе	ОН	ОН	ОМе	Guatteria discolor
Discretine	OMe	ОН	OMe	ОМе	Duguetia obovata
					Guatteria discolor
					G. scandens
					Pachypodanthium staudtii
					Xylopia discreta
Govadine	ОН	OMe	ОН	ОМе	Corydalis govaniana
Govanine	ОН	OMe	OMe	OMe	Chasmanthera dependens
					Corydalis govaniana

Xylopinine	OMe	OMe	OMe	OMe	Duguetia obovata
					Guatteria scandens
					Polyalthia oligosperma
					Stephania suberosa
					Xylopia buxifolia
					X. discreta

3.8 Tetrahydroprotoberberines methylated at C-8 or C-13

Nama		Plants					
Name	2	2 3 8 9				13	Trans
Apocavidine	ОН	OMe	-	-0-C	H ₂ -0-	Me	Corydalis tuberosa
Cavidine	OMe	OMe	-	-0-0	CH ₂ -0-	Me	Corydalis thalictrifolia

Table 3 (continue)

Corybulbine	OMe	ОН	-	ОМе	OMe	Me	Corydalis ambigua C. platycarpa C. tuberosa
Corydaline	ОМе	OMe		ОМе	OMe	Ме	Corydalis ambigua C. aurea C. platycarpa C. tuberosa
Corydalidzine	OMe	ОН	-	OMe	ОН	Me	Corydalis koidzumiana
Epiapocavidine	-0-C	H ₂ -0-	-	OMe	ОН	Me	Corydalis tuberosa
Isocorybulbine	ОН	OMe	-	OMe	ОМе	Ме	Corydalis cava
Lienkonine	OMe	OMe	Me	ОН	OMe	-	Corydalis ochotensis
Tetrahydro- corysamine	-0-C	H ₂ -0-	-	-0-0	Н2-0-	Ме	Corydalis cava
Thalictricavine	-0-0	CH ₂ -0-	-	OMe	OMe	Me	Corydalis tuberosa

Thalictrifoline (Thalictrofoline)	OMe	OMe	-	-O-CH ₂ -O		-0-CH ₂ -0		Me	Corydalis thalictrifolia
Yuanhumine	OMe	OMe	-	OMe	ОН	Ме	Corydalis turtschaninovii		

3.9 Tetrahydropseudoprotoberberines methylated at C-8

Name		St	ructu	re	Plants	
	2	3	8	10	11	Plants
Corytenchirine	OMe	0Me	Me	ОМе	ОН	Corydalis ochotensis

3.10 Tetrahydroprotoberberine N-methylated

Name	,	Stru	cture	4	Plants
Name	2	3	9	10	Trants
Cyclanoline (Cissamine)	ОН	OMe	ОН	ОМе	Cissampelos pareira Stephania tetrandra
Escholidine	-0-С	H ₂ -0-	OMe	OH	Eschscholtzia californica E. douglasi E. glauca Hunnemania fumariaefolia
N-Methylcorydalmine	OMe	OMe	OMe	ОН	Stephania elegans

N-Methyl- isocorypalminium	ОН	OMe	OMe	OMe	Glaucium squamigerum
N-Methylsinactine	OMe	OMe	-0-C	H ₂ -0-	Fumaria officinalis
Steponine	OMe	ОН	ОН	OMe	Stephania japonica

3.11 Tetrahydropseudoprotoberberines N-methylated

Name		Strue	cture		Plants		
	2	3	10	11			
Phellodendrine	ОН	ОМе	OMe	ОН	Phellodendron amurense		

3.12 Tetrahydroprotoberberines hydroxylated at C-13

V		Str	uctur	e						
Name	2	3	9	10	13	Plants				
13-Hydroxytetra- hydropalmatine	OMe	ОМе	ОМе	OMe	ОН	Corydalis ophiocarpa				
13-Hydroxystylopine	-0-C	Н2-0-	-0-C	H ₂ -0-	ОН	Corydalis ophiocarpa				
Ophiocarpine	-0-C	H ₂ -0-	OMe	ОМе	ОН	Cocculus pendulus Corydalis campulicarpa C. cheilanthifolia C. govaniana C. ophiocarpa				

3.13 Tetrahydroprotoberberines with uncommon oxygenation patterns on ring A and ring D

Name			Str	Plants				
	1	2	3	4	9	10	11	110105
Capauridine	ОН	OMe	ОМе	-	ОМе	ОМе		Corydalis aurea
(dl-Capaurine)								C. cava
								C. micrantha
								C. montana
								C. ophiocarpa
								C. pallida
								Stephania glabra
								S. kwansiensis

Table 3 (continue)

Capaurimine	ОН	OMe	OMe	-	OMe	ОН	-	Corydalis montata C. pallida Stephania suberosa
Caseanadine	ОН	OMe	-	-	OMe	OMe	_	Corydalis caseana
Clarkeanidine	ОН	OMe	-	-	ОН	OMe	-	Corydalis clarkei
O-methylthaicanine	-	OMe	OMe	OMe	OMe	OMe	-	Parabaena sagittata
Stepharotine	-	OMe	OMe		OMe	OMe	ОН	Stephania rotunda
Thaicamine	_	OMe	ОМе	ОН	OMe	OMe	-	Parabaena sagittata

3.14 Tetrahydropseudoprotoberberines with uncommon oxygenation patterns on ring A

Name		St	ructu	re	Plants		
	1	2	3	10	11	1101100	
Caseadine	ОМе	ОН	-	OMe	OMe	Corydalis caseana	
Caseamine	ОН	OMe	-	ОН	ОМе	Corydalis caseana	
Stephabinamine	ОН	0Me	OMe	OMe	ОН	Stephania suberosa	
Tetrahydrostephabine	ОН	ОМе	OMe	ОМе	OMe	Stephania suberosa	

3.15 Dihydroprotoberberines

		Stru	cture		
Name	2	3	9	10	Plants
Dihydropalmatine	0Me	ОМе	ОМе	OMe	Stephania kwansiensis
Lambertine	-o-c	Н2-0-	ОМе	OMe	Berberis lambertii

	_		
3.16	Proto	herherine	s N-oxides

Name	Structure	Plants
Carpoxidine		Corydalis ophiocarpa
(Ophiocarpine		
N-oxide)	O N+ NO	
	HO OCH3	
Corynoxidine		Corydalis koidzumiana
	H ₃ CO	
	H ₃ CO H OCH ₃	
	OCH3	

Table 3 (continue)

Nokoensine	H ₃ CO	Corydalis nokoensis
Xylopinine N-oxide	H ₃ CO H ₃ CO H OCH ₃	Stephania suberosa

3.17 Retroprotoberberines

Name	Structure	Plants
lborine	OCH ₃ HOH ₂ C OCH ₃ OCH ₃	Papaver alboroseum P. nudicaule P. oreophilum P. pseudocanescen P. pseudo-orientale P. pyrenaicum
Dehydroorientalidine	OCH3 N+ OCO OCO	Papaver albaroseum P. nudicaule P. areophilum P. orientale P. pseudocanescen P. Pyrenaicum

Aryapavine		Papaver pseudoorientale
	HOH ₂ C OCH ₃	
Mecambridine (Oreophiline)	✓°	Meconopsis cambrica Papaver spp.
	OCH ₃ OCH ₃ OCH ₃	
Orientalidine	OCH 3 N	Papaver bracteatum P. orientale P. pseudo-orientale
	OCH3	

Name		Structure	Plants		
sjatrorrhizine			Jatrorhiza palmata		
		OCH ₃			
		OCH ₃			
		H 3CO N+			
		HO			
		H3CO N+			
		OCH3			
		OCH3			
	4				

0,0-Diacetylcoreximine		Guatteria ouregou
	H ₃ CO	
8-Hydroxyberberine (Berberinol)	OCH3	Arcangelisia flava
11-Hydroxy-12- methoxycoptisine	H ₃ CO OH O	Coptis groenlandica

13-3-Hydroxy-N- methylstylopinium	HO N+CH3	Papaver atlanticum
Karachine	OCH3	Berberis aristata
13-Methoxyoxoberberine	H ₃ CO CH ₃	Berberis darwinii

0-Methylprechilenine		Berberis darwinii
	OCH ₃	
Oxotetrahydropalmatine		Anamirta cocculus
	$_{\rm H_3CO}$ $_{\rm OCH_3}$	
Oxyberberine (Berlambine)	OCH3 OCH3	Berberis empetrifolia B. lambertii B. oblonga Thalictrum foliolosum T. longistylum T. minus T. podocarpum

8-Oxypseudopalmatine		Stephania suberosa
	$\begin{array}{c} {\rm H_3CO} \\ {\rm H_3CO} \\ \\ \\ {\rm OCH_3} \end{array}$	
Prepseudopalmanine		Berberis darwinii
	H_3^{CO} H_3^{CO} H_0 OCH_3	
Solidaline	H ₃ C0	Corydalis solida
	H ₃ CO OCH ₃ OCH ₃	

Staudine	H ₃ CO H ₃ CO OCH ₃ OCH ₃	Pachypodanthium staudtii
Stephabine	H_3CO OH N^+ OCH_3	Stephania suberosa
Valachine	OCH3	Berberis valdiviana

2.3 Oxidation and reduction of protoberberine alkaloids

The dihydroprotoberberines (d) and tetrahydroprotoberberines (b) could be oxidized to the corresponding quaternary protoberberines(a) with iodine, mercuric acetate, or simply by standing in air (scheme 1). The quaternary protoberberines (a) could be reduced to the parent tetrahydroprotoberberines (b) with a variety of reducing agents such as mixed metal hydride, zinc/hydrochloric acid, and catalytic reduction in the presence of platinum. If, however, the reduction is carried out with a mixed hydride in a dry aprotic solvent, the reaction stops at the dihydroprotoberberine (d) stage. The quaternary protoberberine salts (a) unstable in the presence of concentrated alkali. They form the oxo derivatives (f) and the dihydro derivatives (d) by hydrogen transfer. The oxo derivatives(f) are exclusively formed from the quaternary protoberberine salt (a) by oxidation with potassium ferricyanide. The dihydroprotoberberine derivative (d) remains in equilibrium with its immonium form (g) in a protic medium. The immonium form (g) is unstable and undergoes rapid disproportionation to a mixture of the quaternary protoberberine salt (a) and tetrahydroprotoberberine(b). (Bhakuni and Jain, 1989).

2.4 Stereochemistry of protoberberine alkaloids

The stereochemistry of alkaloids such as ophiocarpine and 13-epi-ophiocarpine can be deduced from their IR spectra. Both compounds display Bohlmann bands at about 2800 cm⁻¹, indicating a trans-fused B/C ring juncture, but only ophiocarpine has a hydrogen-bonded hydroxyl group. This indicates internal proximity of the hydroxyl proton and the nitrogen lone pair; a situation possible only when the 13-hydroxy group is beta and axial as in ophiocarpine.

Ophiocarpine

13-Epi-ophiocarpine

These very simple concepts do not hold however when substituents are at position 1, 8 or 13 (other than OH). In particular a compound may show Bohlman bands in a KBr pellet, but not in solution, indicating subtle conformational inversion, trans to cis, at the nitrogen between crystalline states and liquid. Such a phenomenon is particularly true of 1-substituted protoberberines.

Junction stereochemistry is through kinetic studies of the rates of methylation of the tertiary nitrogen with methyliodide. In general terms the cis-quinolizidine compounds react much faster than the corresponding trans isomerchanging the substitution also changes the basicity of the nitrogen so that 10,11-substituted isomers show faster rates of N-methylation than 9,10-substituted isomers. When substitution is by a phenolic group, the rates of methylation are between those expected for cis- and transquinolizidines and consequently such data must be used with considerable caution.

An interesting demonstration of the differences in stereochemisty of two 13-methyl-substituted tetrahydro-protoberbeines is observed on Hofmann degradation to afford quite different products depending on the availability of the β -hydrogen trans to the nitrogen lone pair. Corydaline for example afforded (a), but mesocorydaline gave the vinyl derivative (b).

The absolute configuration of the protoberberines was deduced when (-)-N-norlaudanasoline of known absolute configuration afforded (-)norcoralydine. The levorotatory bases have the same absolute configuration as (-) norcoralydine and also exhibit a negative ORD curve in the region of 240 nm. (Cordell, 1981)

2.5 Aporphine alkaloids

The aporphines are the largest group of isoquinoline alkaloids and are represented by the general structure below.

General structure of aporphine alkaloids

These alkaloids are distributed in at least 18 plant families, of which the most important are the Papaveraceae, Anonaceae, Lauraceae, and Monimiaceae. The nitrogen atom is usually methylated, making that nitrogen tertiary. If the nitrogen is secondary, the alkaloid is called a noraporphine, they are not very stable and are often characterized as their N-acetyl derivatives. Several quaternary aporphine salts with two methyl groups attached to the nitrogen are also known. Aporphines are known with the C-6a stereochemistry either or .

The most diverse structural feature of the aporphines is the oxygenation pattern. Positions 1 and 2 are always oxygenated, either by hydroxy, methoxy or methylene dioxy

groups. It is common to find further oxygen substituents at C-9, C-10 and C-11, and occasionally at C-8. It is rare to find oxygenation at C-7, except in the oxoaporphines, and even rarer to find any oxygenation in ring B.(Cordell, 1981)

The first aporphine alkaloid was obtained not by isolation but as the result of chemical reaction. Thus in 1969 it was found that hot concentrated hydrochloric acid caused rearrangement of morphine (M) to apomorphine (Ma) which is not a natural product. (Cordell, 1981)

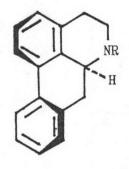
Morphine(M)

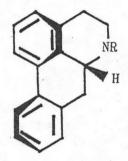
Apomorphine (Ma)

Since that time, nearly 200 natural aporphine alkaloids have been recognized. All of these incorporate in tetracyclic nucleus present in apomorphine. Additionally, they are without exception oxygenated at C-1 and C-2, and very often at other positions as well. The oxygenated substituents are usually hydroxy, methoxy or methylenedioxy groups. (Shamma and Guinaudeau, 1984)

2.6 Stereochemistry and absolute configuration of the apophines

The stereochemistry and absolute configuration of the aporphines have been thoroughly studied. Shamma in 1960 was first to make an interesting observation concerning the lack of planarity of the biphenyl system, and indeed two stereochemical possibilities exist as shown below: (Cordell, 1981)





L-S configuration

D-R configuration

Bentley and Cardwell first observed that since naturally occurring (+) -glaucine, and (-) -morphothebaine, a rearrangement product from codeine are enantiomeric at C-6a and the absolute configuration of (-) -morphothebaine is known to be of the D series, (+) glaucine must belong to the L configuration. They also generalized that all aporphines that are appreciably dextrorotatory at the sodium D line belong to the L series, whereas the levorotatory aporphines are of the D configuration. (Shamma, 1972)

The absolute configuration of natural (+)bulbocapnine is also known with certainty and first the above rotation rule since Ayer and Taylor have converted the alkaloid by treatment with sodium in liquid ammonia into (+)morphothebaine of known absolute configuration.(scheme 2) Additionally, a three dimensional x-ray analysis of bulbocapnine methiodide has been carried out. The angle of twist of the biphenyl system is 29.9. The distance between 0-1 and 0-11 is 2.74 °A which is just under the sum of the van der Waals radii of the two oxygens, which is 2.80 °A this data is example that the biphenyl system in bulbocapnine methiodide is appreciably strained. (Shamma, 1972).

(+) -Bulbocapine (+) -Morphothebaine

Scheme 2 Absolute configuration of bulbocapnine

The ORD curves of aporphines have been studied. Craig and Roy have noted that aporphines exhibit a cotton effect of high amplitude centered between 235 and 245 mm. This curve is independent of the substitution at the 1, 2,3, 9, 10 and 11 positions and is diagnostic of the absolute configuration; if the cotton effect is positive, the alkaloid belongs to the L series, and if the cotton effect is negative, the compound must be of the D series.

Simple specific rotations at the sodium D line, besides being fairly reliable indicators of the absolute configuration, offer a simple way of differentiating between C-1, 2, 9, 10- and C1, 2, 10, 11 -substituted aporphines. The 1, 2, 9, 10- series exhibits rotations of +119 or less, whereas the 1, 2, 10, 11 -substituted aporphines show values of +139 or more. The occurrence of racemic aporphines in nature is rare. (Shamma 1972)

2.7 Alkaloids isolated from Coscinieae tribe

Over the past 160 years ago alkaloids have these plants recieved considerable attention. The major alkaloids, which have been isolated are protoberberine type, and the main alkaloid is berberine. The more recent work has not resulted in any unexpected novel structure of alkaloids but rather in the isolation of new isomers together with observation of alkaloidal pattern variation. The alkaloids which have been reported in these 5 species of Coscinieae are shown as follows:-

2.7.1 Alkaloids from Anamirta cocculus (L.) Wight et Arn.

Anamirta cocculus (L) Wight et Arn. is a liana which occurs in southeast Asia. Berries of this plant are used as a fish poison. From the berries the sesquiterpene mixture picrotoxin is commercially isolated. The berries have been official in a number of pharmacopaeias. The seed shells of this plant were reported to contain alkaloids in 1834 by Pelletier and Couerbe, the two alkaloids isolated, were menispermine $(C_{18}H_{24}O_{2}N)$ and paramenispermine $(C_{18}H_{24}C_{2}N)$. No other reports on alkaloids in Anamirta cocculus (L) Wight et Arn. have been published since the work of Pelletier and Couerbe. Until, in 1980, the studies on Indonesia medicinal plants was reported

(Siwon, Verpoorte, Tieken and Svenden, 1980). In the stems and roots of Anamirta cocculus (L.) Wight et Arn., the quaternary protoberberine alkaloids berberine, palmatine, columbamine and L-8-oxotetrahydropalmatine were isolated. There is one of isoquinoline alkaliod was isolated too, magnoflorine.

The occurrence of protoberberine-type alkaliods in Anamirta cocculus (L.) Wight et Arn. is not surprising. The genus Anamirta belongs to the tribe Coscinieae of the Menispermaceae. The other two genera in this tribe. Arcangelisia and Coscinium, also contain protoberberine alkaloids. Particulary Arcangelisia is botanically closely related to Anamirta, and Arcangelisia flava (L.) Merr. and Anamirta cocculus (L.) wight et Arn. have sometimes been confused.

2.7.2 Alkaloids from Arcangelisia species

The genus Arcangelisia belongs, together with the genera Coscinium and Arcangelisia, to the tribe Coscinieae of the Menispermaceae. The genus Arcangelisia comprises of two species, A. flava (L.) Merr. is a liana found in Southeast Asia, A. tympanopoda(Lauberb & K. Schum.) Diels. is a liana found so far only in New-Guinea. The two species are differentiated by the size of the fruits, the latter one having larger fruits than the former.

Various medicinal uses of A. flava (L.) Merr. have been reported as a febrifuge, tonic, abortive, expectorant and against hepatitis and digestion. Since 1931, Santos repoted the presence of berberine in the stems of A. flava (L.) Merr. Santos identified berberine, jatrorrhizine and columbamine and isolated an alkaloid which was called "shobakunine". Thes was later shown to be a mixture of palmatine and berberine.

Following this quite an amount of work was carried out on A. flava (L.) Merr. from Philippines by van Steenis Kondo, Estrada et al. They reported only berberine, jatrorrhizine and columbamine. In 1988 Garcia et al detected berberine, palmatine and jatrorrhizine in the stems and berberine and jatrorrhizine in the roots of A. laureirii Diels. (a synomym for A. flava) (Garcia, 1881). At that time the studies of Indonesian medicinal plants VII by Verpoorte, et al, isolated and identified six quaternary alkaloids, thalifendine, dehydrocorydalmine, jatrorrhizine, pycnarrhine, berberine and palmatine, and three tertiary alkaloids, hydroxy-berberine, limacine and homoaromoline form stems and roots of Arcangelisia flava (L.) Merr. (Siwon et al, 1982). For Arcangelisia tympanopoda (Lauberb & K. Schum.) Diels, however, it is notable that there has no phytochemical report about it.

2.7.3 The alkaloids of Coscinium species

The genus Coscinium consists of two species only the widely distributed C.fenestratum (Gaetn.) Colebr. (C. wallichianum Miers or C. usitratum Pierre) and C. blumeanum Miers, known only from Peninsular Thailand and the islands off the west coast of Malaya.

Berberine, palmatine and jatrorrhizine have been isolated from roots and stems of *C. wallichianum* (*C. fenestratum*) by Garcia *et al* in 1969 (Garcia, *et al*, 1970). In 1981 Siwon and Verpoorte have collected *Coscinium fenestratum* (Gaertn.) Colebr. on south Kalimantan, Indonesia and major alkaloids berberine and jatrorrhizine have been isolated. Apperciable amounts of berberubine and N,N-dimethyllindcarpine and small amounts of thalifendine and palmatine are also present. Stems and roots contain the same pattern of alkaloids.

For *C. fenestratum* (Gaertn.) Colebr. which collected from India in 1988, apart from berberine, oxyberberine, tetrahydroberberine (canadine), a new minor alkaloid 12,13-dihydro-8-oxo-berberine have been furnished by Malhotra, Taneja and Dhar. (Malhotra et al, 1989).

Tomita and Tani isolated palmatine, berberine and jatrorrhizine from Coscinium blumeanum Miers collected in Sararvak (North Kalimantan, Malaysia) in 1940.

The presence of protoberberine alkaloids in roots and stems of Coscinieae tribe is consistent with their uses in curing microbial infection in folk medicine.

3. Biosynthesis

3.1 Biogenesis of protoberberine alkaloids

There are two terms, biosynthesis and biogenesis, commonly used in discussing the formation of secondary metabolism products. Biosynthesis is the experimental study of the formation of secondary metabolites. Whereas biogenesis is the hypothetical speculation on the precursor-product relationships in a biosynthetic pathway. By means of this definition biogenesis refers to the manner in which the organic substances are synthesized, altered, or degraded by plant or animal organisms. It is based mainly on the visual dissection of a molecule into recognizable precursor fragments. (Cordell, 1981)

The biogenesis of protoberberine alkaloids was discussed earlier. The relationship of 1-benzyltetrahydro-isoquinolines and protoberberines was recognized quite early. As a result of numerous studies, the biosynthesis of the berberine alkaloids has been well worked out and the currently accepted scheme is shown in scheme 3.

Scheme 3 Biosynthesis of the protoberberine alkaloids

The fundamental units are those for the formation of the benzylisoquinoline alkaloids with the addition of a single carbon atom which becomes C-8 of the skeleton. Thus two molecules of tyrosine are involved, one proceeding to dopamine via dopa, and the second to 3,4-dihydroxyphenyl pyruric acid.

Tyrosine

Dopa

Dopamine

$$\mathsf{HO} \overset{\mathsf{CO_2H}}{\longrightarrow} \mathsf{O}$$

4-Hydroxy phenyl pyruvic acid

3,4-Dihydroxy
phenyl pyruvic acid

Thus $[2^{-14}c]$ tyrosine labels at C-6 and C-13 of berberine in *Hydrastis canadensis L.* (Ranunculaceae), but $[1^{-14}c]$ dopamine labels only at C-6. Shown below:

However, later observation in several tracer experiments, reticuline was shown to be a biological precursor of protoberberine alkaloids. The early stages of biosynthesis of these alkaloids from reticuline was studied.

The C-8 atom of protoberberine and derivative alkaloids is known as the "Berberine bridge". It has been suggested that the bridge could be formed in nature by oxidative modification of an N-methyl group. Plausible machanisms of the reaction are shown in scheme 4.

$$\begin{array}{c} & & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & &$$

Reticuline

OH

осн3

$$(C)$$

$$(d)$$

$$(d)$$

$$(d)$$

$$(d)$$

$$(d)$$

Scheme 4 Mechanism of berberine bridge formation

Two electron oxidation of a suitable precursor such as reticuline could give the phenoxonium ion (a) from which (b) could be derived by intramolecular hydrogen transfer. In an alternative mechanism, one electron oxidation of reticuline could furnish a phenolate radical. A hydrogen atom could then be transferred from the N-methyl group to oxygen, which could be oxidized to the biradical (d). Coupling of radicals could then lead to ring closure to give (e) as in normal c-c bond formation in phenolate oxidaton reactions. Experimental support in favor of the hypothesis was put forward by Barton, Battersby and co-workers directly and by Gupta and Spenser indirectly. Barton et al fed reticuline labeled with (^{14}c) in its N-methyl group to Hydrastis canadensis. Biosynthetically synthesized berberine was degraded unambiguously and essentially all of the radioactivity was found at C-8 of the alkaloid. Battersby's group by using laudanosoline labeled with [14c] in the methyl group had confirmed the above results. Gupta and Spenser had provided evidence from experiments with methionine -14C.

Barton and co-workers have studied biosynthesis of berberine in detail. Feeding doubly labeled reticuline, they confirmed that C-8 of berberine was derived from the N-methyl group of the precursos. Further, it was demonstrated that the methylenedioxy group in berberine was also formed as in other cases by oxidative cyclization of a catechol -O- methyl ether. Parallel feeding of both the labeled enantiomers of reticuline showed that (+)

reticuline was converted to berberine 15 times more efficiency than (-) reticuline in *H. canadensis* plants (scheme 5) (-)canadine occurs in *H. canadensis*. Efficient incorporation (8.9%) of (+) canadine into berberine by the plants established its intermediacy, protosinomenine, however, was not incorporated into berberine in the plants. (Bhakuni and Jain, 1989)

Berberine

Canadine

Feeding label (+) laudanosoline to berberis japonica Lindl. (Berberidaceae) also gave rise to labeled berberine. Labeled (+) reticuline significantly incorporated into (-) stylopine in Chelidonium majus L. (Papaveraceae), while incorporation of (-) reticuline is a much less efficient process. Some of the tritium label at C-14 in (-) stylopine was lost, indicating C-1 oxidation-reduction at the (+) reticuline stage, shown below:

The oxidation of the N-methyl group and subsequent ring closure is thought to proceed *via* the iminium species to give either the 9,10 - (pathway a) or 10,11-disubstituted (pathway b) series of compounds. (Cordell, 1981)

The formation of the methylenedioxy group from an o-methoxyphenol can be visualized either in terms of an ionic or a free-radical machanism, and the former is depicted here. scheme 6.

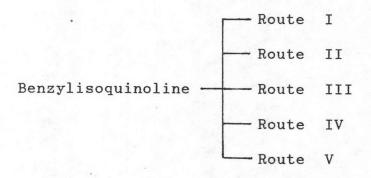
Scheme 6 The formation of methylenedioxy

Turning to the 5-hydroxylated protoberberine alkaloids, berberastine, thalidastine and tetrahydroberberastine, it has been shown that dopamine and noradrenaline are incorporated into berberastine much more efficiently than into canadine or berberine. The last two alkaloids, therefore, cannot be precursors for berberastine. Rather, the C-5 hydroxyl must be introduced at some early stage which preceeds the formation of the necessary tetrahydrobenzylisoquinoline intermediate. (Shamma, 1972).

The protoberberines are not only interesting in themselves from a biosynthetic point of view, but are important because they act as precursors to other skeleta. In particular, these alkaloids (e.g. scoulerine) are the precursors of the protopine, rhoeadine, benzophenanthridine (e.g.chelidonine), and phthalideisoquinoline (e.g.narcotine) In addition they are thought to be the precursors of the spirobenzylisoquinoline alkaloids (e.g. ochotensamine) (Cordell, 1981) shown in scheme 7.

3.2 Biosynthesis of aporphine alkaloids

The discussion on the biogenetic synthesis of the aporphine alkaloids has raised a number of points concerning the biosynthesis of this group. In particular and depending on the orientation of phenolic and methoxy groups might envisage any of at least five routes being in operation from a benzylisoquinoline precursor (Figure 1). In vitro all these routes appearently can operate.



Route I → proaporphine → aporphine

Route II→neoproaporphine → aporphine

Route III-morphinandienone → neoproaporphine aporphine

Route IV-directly coupled 3a-substituted quinol → aporphine

Route V - direct coupling of radicals -> aporphine

Figure 1 Biogenesis of aporphine from benzylisoquinoline

The structure of the aporphine alkaloids (+) roemerine and (+) -isothebaine indicated that they
are probably not derived from a tetrahydrobenzylisoquinoline
by direct coupling.

Barton and co-workers investigated the formation of (+)-roemerine in *Papaver dubium* L. and found that tritium labeled (+) -N- methylcoclaurine was well incorporated. The position of the hydroxy group in the precursor suggests that a proaporphine intermediate is involved (scheme 8) similarly.

Scheme 8 Proaporphine intermediate is involved

Battersby and co-workers showed that (+)-orientaline was well incorporated into (+) - isothebaine in *Papaver orientalis*. The formation of (+)isothebaine can be envisaged as occurring through (-)-orientalinone, a co-constituent, as shown in scheme 9.

$$H_3CO$$
 H_3CO
 H_3C

Scheme 9 The incorporated from orientaline to isothebaine

(+) -isothebaine

Brachmam & Haussen found that [N-methyl-14C] reticuline was a precursor of isoboldine, but not of magnoflarine in *Papaver somniferum*. Classically, these would be regarded as the products of ortho-para and ortho-ortho coupling of reticuline. Subsequently the same group showed that [N-methyl-14C] reticuline was a precursor of magnoflerine in *Aquilegia* sp. shown below.

The biosynthesis of boldine in Litsea glutinosa (Lour.) C.B.Rob.var. glabraria Hook. (Lauraceae) has also been studied in detail by Kapil and co-workers, with a quite different result.

Of several variously methylated benzylisoquinoline precursors only 4 -O-methylnorlaudanosaline, nor-reticuline, and reticuline were precursors. In the latter case the (+)isomer reticuline was in corporated with considerable preference into boldine. More surprising however is the high (2%) level of incorporation of [8- 3 H] isoboldine into boldine, a process that of necessity involves 2- 3 C-demethylation followed by 1- 3 C-methylation. (scheme 10)

Scheme 10 Biosynthesis of boldine in Litsea glutinosa var. glabaria

Norprotosinomenine was not a precursor of boldine in this plant, which contrasts with the previously discussed work with boldine in *Dicentra eximia*.

More definitive results have been obtained by Battersby and co-workers concerning the formation of corydine and glaucine in Dicentra eximia (Ker.) Torr. (Fumariaceae). Reticuline and orientaline were not precursor, but 4"-O-methylnorlaudanosoline and norprotosinomenine were effective precursors. This result clearly rules out direct phenol coupling (for corydine and reticuline) and a proaporphine intermediate. According to Battersby this suggests the pathway shown in scheme 11 involving two alternative neoproaporphine intermediates (A) and (B). Notice that the aporphine (C) has the incorrect omethylation pattern in comparison with corydine. It would be interesting indeed to know if 1-O-demethylation followed by 2-O-methylation really occurs at this point in the biosynthesis. Boldine was a poor precursor or glaucine.

Scheme 10 Biosynthesis of boldine in Litsea glutinosa var. glabaria

These results would suggest that boldine is produced by two different pathways in two different plants. If this is a general trend, the biosynthesis of aporphine alkaloids may never be established. Clearly this is an area in need of considerable further study, and at this time it is not clear which, or how many, of the possible biosynthetic routes may be operating in order to produce the various aporphine alkaloids. (Cordell, 1981)

4 Pharmacology

4.1 Pharmacological activity of protoberberine alkaloids

Protoberberine alkaloids and their derivatives exhibit several types of biological activities. However, to date berberine alone was found to be of clinical value and is being used in the treatment of gastrointestinal disorder.

of the most important actions is antimicrobial activity of berberine and derivatives, which covers the range of organisms from fungi and protozoa to bacteria. In 1947, Sado has been examined systematically antibacterial activity of berberine chloride, iodide, and palmatine iodide against Vibrio, Eberthella, Salmonella and Escherichia organisms and observed that the antibacterial action of berberine chloride was virtually invariant between pH 5 and 9. Berberine sulfate inhibited the growth of Candida tropicalis and Xanthomones citri at a concentration of 3.1 Mg/ml and of Pseudomonas and Salmonella at a concentration > 100 µg/ml. Lahiri and Dutta have recommended the use of berberine as an adjunct to isotonic saline and electrolyte replacement therapy in acute cholera (Lahiri and Dutta, 1967). Berberine is reported to depress intestinal peristalsis and to remove inflammatory congestion of the mucosal surface of the intestine. It is also effective the treatment of diarrhea of infancy and childhood. Berberine has been shown to form a complex with DNA, probably intercalating into supercoiled mitrocondrial DNA to produce configurational change in DNA (Shamma, 1972)

Better antibacterial activity than berberine has been demonstrated by salts of berberine and sulfanilamide. However, appropriate comparisons with standard antibacterial are lacking. Berberine chloride is found to eliminate. Syphacia obvelata from the intestine of mice. Berberine is also found effective in the treatment of cutaneous leshmaniasis. Antifungal and antiarrhythmic activities have been shown by a number of 8 ß-substituted berberines. Berberine sulfate and tetrahydropalmatine inhibit the respiratory chain by interfering with the action of NADH oxidase berberubine chloride isolated from Thalictrum polyganum is found to prosses antimicrobial activity against Mycrobacterium smegmatics at 100 µg/ml. (Bhakuni and Jain, 1989)

The anticancer activity of protoberberine have been reviewed by Suffness and Cordell (The alkaloids, 1985). Berberine has been reported to prossess cytotoxic and neoplasm inhibitory activity against KB and Ehrlich ascite tumor cell, but the derivative coralyne chloride displays activity against both the P-388 and L-1210 lymphocytic leukemia systems in mice. A number of coralyne salts and analogs have been synthesized, and structure activity relationship has been studied. Although different salts of coralyne have comparable activity, coralyne acetosulfate is found most active. The planarity and regidity of molecules of these types are found critical activity. Coralyne formed a stable complex with thymus DNA in vitro, which is found

responsible for its activity.

Contractive activities of tertiary and quaternary berberine-type alkaloids have been studied on isolated uteri of mice. Quaternary protoberberines including berberine, palmatine, jatrorrhizine, coptisine, and dehydrocorydaline cause marked contraction of uterine muscles, but only a weak spasmolytic activity is exhibited on isolated intestines of mice. On the contrary berberines including canadine, tetrahydropalmatine and tetrahydrojatrorrhizine show strong papaverine-like action, although their contractive activities on uteri are transitory dehydrocorydaline chloride is found to prossess considerable gastric antisecretory activity. When administered orally to rat and guinea pigs, it prevents gastric and duodenal ulcers.

Pavelka and Kovar (Pavelka and Kovar, 1975) have studied the liver alcohol dehydrogenase activity of several protoberberine alkaloids. 13-Ethylberberine has been found to be the most active inhibitor of liver alcohol dehydrogenase the compound is bound more firmly to the enzyme at pH 10 than NAD and NADH.

The quaternary protoberberine alkalaids, berberine, coptisine, and substituted berbine are found to be weak inhibitors of butyrylcholinesterase in human serum, whereas jatrorrhizine and columbamine are found to be more potent.

The pharmacological actions and effects reported in the studies of protoberberine alkaloids, both of the natural (marked with asterisk) and synthetic compounds, are listed in table 4 according to the classification (Simeon et al, 1989)

Table 4 Pharmacological activities of protoberberine alkaloids

The simple 5,6-dehydropseudoprotoberberines

Name	2	3	8	9	10	11	12	Activity
Dehydroberberubine	-0-CH ₂ -0-		-	ОН	OMe	-	-	Antitumoral
Isocoralyne	ОМе	OMe	Me	-	-	OMe	OMe	Antileukemic

Quaternary protoberberines

Table 4 (continue)

	\$	Struct	ure		
Name .	2	3	9	10	Activity
9-Acetylberberrubine	-0-CH ₂ -0-		-O-CH ₂ -O- OAc OM		Antitumoral Ganglionic stimulant block Parasympathomimetic
9-Benzoylberberubine	-O-CH ₂ -O-		OBen	OMe	Antitumoral
Berberine*	-0-CH ₂ -0-		OMe	OMe	<pre></pre>

Table 4 (continue)

Berberine*	Antitumoral
	Antiulcer
	Blocking R factors
	Cholagogue
	Cholecystokinetic
	Choler toxin antagonist
	Choleretic
	CNS depressant
	Competitive adrenalytic
	Cytotoxic
	Decrease blood level of
	cholesterol
	Decrease blood level of T
	Decrease Ca ⁺² available in
	intracellular receptors
	Decrease urinaty volume
	Ganglionic stimulant bloc
	Hypoglycemic
	Hypothermia
	Increase bilirubin
	excretion
	Increase lachrymal
	secretion

Table 4 (continue)

		 		r
Berberine*				Increase the action of
berberine				
	314			antitumorals
				Increase the activity of
				the UDP-glucuronyl
				transferase
	**			Inhibition of
				Inhibition of
				acetylcholinesterase
				Inhibition of alcohol
				dehyddrogenase .
				Inhibition of aldehyde
				reductase I
				Inhibition of cation-
				dependent
				ATP-phosphohydrolase
				Inhibition of diamine
				oxidase
				Inhibition of lactic
				fermentation
				Inhibition of NADH oxidas
				Inhibition of reverse
				transcriptase
				Inhibition of RNA
				synthesis
				Inhibition of
			200	tryptophanase

Table 4 (continue)

Berberine*		Inhibition of tyrosine
		decarboxylase
		Inhibits enterotoxins
		Inhibits the action of
		carcinogens
		Inhibits the synthesis
		of DNA
		Intestinal antisecretory
	13.	Local anesthetic
		Mutagenic
		Mydriatic
		Negative chronotropic
		Parasympatholytic
		Parasympathomimetic
		Phototoxic
		Pilomotor erection
		Positive chronotropic
		Positive inotropic
		Ptosis
		Sedative
		Spasmolytic
		Spermicide
		Sympatholytic
		Uterotonic
		Vasodilator

Table 4 (continue)

n *	0.00		OII	OM	Antinionalial
Berberrubine * (9-Berberobine)	-0-CH	12-0-	ОН	OMe	Antimicrobial Antitumoral Ganglionic stimulant block Hypotensive Negative chronotropic Parasympatholytic Parasympathomimetic
O-Butylberberrubine	-O-CH ₂ -O-		OBu	OMe	Hypotensive Inhibition of cardiac function Negative chronotropic Parasympatholytic
Columbamine *	ОН	OMe	OMe	OMe	Antimicrobial Inhibition of butyrylcholinesterase Uterotonic
Coptisine *	-0-CH ₂ -0-		-0-CH ₂ -0-		Anti-inflammatory Antimicrobial Cytotoxic Inhibition of acetylcholinesterase

Table 4 (continue)

Coptisine*					Inhibition of alcohol dehydrogenase Spasmolytic Uterotonic
D-Dodecylberberrubine	-O-CH	12-0-	OD	OMe	Hypotensive
Jatrorrhizine *	OMe	ОН	OMe	OMe	Antiarrhythmic Anti-inflammatory Antimicrobial Inhibition of butyrylcholinesterase Spasmolytic Uterotonic
0-Octylberberrubine	-0-0	H ₂ -0-	00Ct	OMe	Hypotensive
Palmatine *	OMe	OMe	OMe	OMe	ACTH-like Anti-inflammatory Antimicrobial Antiparasitic Antipyretic Antitumoral Hypotensive

Table 4 (continue)

Quaternary pseudoprotoberberines

		Struct	ture		
Name	2	3	10	11	Activity
Pseudocoptisine	-O-CH ₂ -O-		-0-СН2-О-		Inhibition of acetylcholinesterase
Pseudoepiberberine *	OMe	OMe	-0-CH ₂ -0-		Inhibition of acetylcholinesterase

Quaternary protoberberines substituted at carbon 13

		Str	ructu	re		
Name	2	3	9	10	13	Activity
Corysamine *	-0-C	H ₂ -0-	-о-СН2-О-		Me	Inhibition of alcohol dehydrogenase
Dehydrocavidine *	OMe	OMe	-0-0	-0-CH ₂ -0-		Analgesic Anticonvulsant Antimicrobial Increase the levels of hepatic glucogen Inhibits uterine contractions Negative inotropic Spasmolytic

Table 4 (continue)

Dehydrocorydaline *	OMe	OMe	OMe	OMe	Ме	Adrenalytic
						Anticonvulsant
						Antihipoxic
						Antitumoral
						Antiulcer
						Decrease urinary
						excretion of sodium
						Hypotensive
						Increase coronary flow
						Increase hypoxia
						tolerance .
						Increase myocardial
						uptake
						Inhibition of reverse
						transcriptase
						Negative inotropic
						Peristaltic stimulant
						Protective effects on
						myocardial necrosis
						Protects pituitary
						induction
						Sedative .
1 77 32						Spasmolytic
						Uterotonic

Table 4 (continue)

3-Methylberberine	-0-СН	2-0- 0)Me	OMe	Me	Antiarrhythmic Antihipoxic Inhibition of acetylcholinesterase Inhibition of reverse transcriptase
13-Methylberberrubine	-O-CI	H ₂ -0-	ОН	ОМе	Me	Antitumoral
13-Methylpalmatrubine	OMe	OMe	ОН	ОМе	Me	Antitumoral
13-Allylberberrubine	-0-C	-0-CH ₂ -0-		OMe	All	Antiulcer Inhibition of butyrylcholinesterase
8-Benzylberberine		-0-CH ₂ -0- 8=Ben		OMe		Antimicrobial
13-Ethylberberine	-0-0	-0-CH ₂ -0-		OMe	Et	Inhibition of acetylcholinesterase Inhibition of alcohol dehydrogenase
13-Propylberberine	-0-	-CH ₂ -O-	- OMe	OMe	P	r Antidiarrheic Miocardial stabilizer

Table 4 (continue)

Pseudoprotoberberine methylated at carbon 8 or 13

	,	S					
Name	2	3	8	10	11	13	Activity
Dihydrocoralyne	OMe	OMe	Me	OMe	OMe	-	Antileukemic
13-Methylpseudoberberine	-0-0	H ₂ -0-	-	OMe	OMe	Me	Antitumoral Inhibition of reverse transcriptase

7-Dihydroprotoberberines Structure Activity 2 3 9 10 Name -O-CH₂-O- OMe Dihydroberberine OMe Uterotonic Dihydropalmatine * OMe Uterotonic OMe OMe OMe

Tetrahydroprotoberberines

		Struc	cture		
Name	2	3	9	10	Activity
(-) Aequaline * ((-) Discretamine)	OMe	ОН	OMe	ОН	Dopaminergic antagonist
(-) Corypalmine) *((-) Tetrahydrojatrorrhizine)	OMe	ОН	ОМе	ОМе	DA ₂ antagonist Dopaminergic antagonist Spasmolytic
(†) Corypalmine ((±) Tetrahydrojatrorrhizine)	OMe	ОН	OMe	OMe	Monoanimes depletor in brain Spasmolytic Tranquillizer

Table 4 (continue)

+) 2,3-Dihydroxy 9,10- dimethoxy tetrahydroprotoberberine	ОН	ОН	OMe	OMe	Inhibition of dopamine- sensitive adenylate cyclase
-) 2,3-Dihydroxy 9,10- dimethoxy tetrahydroprotoberberine	ОН	ОН	OMe	ОМе	Inhibition of -sensitive adenylate cyclase Inhibition of dopamine- sensitive adenylate cyclase
(*) 2,3-Dihydroxy 9,10,11- trimethoxy tetrahydroprotoberberine	ОН	ОН		OMe 1-OMe	Inhibition of S-sensitive adenylate cyclase
(+) 9,10-Dihydroxy tetarhydroprotoberberine	-	-	ОН	ОН	Inhibition of dopamine sensitive adenylate cyclase
(-) 9,10-Dihydroxy tetrahydroprotoberberine	-	-	OI	H OH	Inhibition of dopamine sensitive adenylate cyclase

Table 4 (continue)

(+) 9,10-Dimethoxy 10- methylthio tetrahydroprotoberberine	OMe	OMe	-	SMe	Tranquillizer
(+) 9,10-Dimethoxy tetrahydroprotoberberine	-	_	ОМе	OMe	Inhibition of dopamine- sensitive adenylate cyclase
(-) 9,10-Dimethoxy tetrahydroprotoberberine	-	_	ОМе	ОМе	Inhibition of dopamine- sensitive adenylate cyclase
<pre>(-) Isocorypalmine * ((-)tetrahydroprotoberberine)</pre>	ОН	OMe	OMe	ОМе	Antitumoral Dopaminergic antagonist
<pre>(-) Kikemanine * ((-) Corydalmine)</pre>	OMe	0Me	ОМе	ОН	Dopaminergic antagonist
Nandinine *	-0-0	CH ₂ -0-	ОН	OMe	Uterotonic
(-) Scoulerine *	ОН	OMe	ОН	OMe	Dopaminergic antagonist
Scoulerine	ОН	OMe	ОН	OMe	Antiemetic Antitussive

Table 4 (continue)

(-) Stepholidine *	ОН	ОМе	OMe	ОН	Analgesic
(-) Stepholidine *	ОН	OMe	OMe	ОН	Antipyretic Antiserotoninic DA2 agonist Dopaminergic antagonist Hypotensive Increase the action of analgesic Inhibition of catecholamine uptake Monoamines depletor in
					brain Sedative Spasmolytic
(±) 2,3,9,10,11-Pentamethoxy tetrahydroprotoberberine	OMe	OMe	OMe	OMe -OMe	Tranquilizer
(+) Tetrahydroberberine *	-0-0	H ₂ -0-	- OMe	OMe	Dopaminergic antagonist Inhibition of dopamine- sensitive adenylate cyclase

Table 4 (continue)

			т —	T
(-) Tetrahydroberberine *	-O-CH ₂ -O-	OMe	OMe	Antipsychotic Clonic spasm Ganglionic stimulant block Hypotensive Inhibition of -sensitive adenylate cyclase Sedative Spasmogenic
(±) Tetrahydroberberine	-O-CH ₂ -O-	OMe	OMe	Analgesic DA ₂ agonist Emetic Hypothermia Monoamines depletor in brain Sedative Spasmolytic
Tetrahydroberberine	-0-СН2-0-	OMe	OMe	Analgesic Antipsychotic Antitussive Ataraxic

Table 4 (continue)

Tetrahydroberberine			CNS depressant
			Curare-like
			Dopaminergic antagonist
			Ganglionic stimulant
			block
			Hypotensive
			Inhibition of
			conditioned reflexes
			Sedative
			Tranquillizer
			Uterotonic
(-) Tetrahydrocoptisine *	-0-CH ₂ -0-	-0-CH ₂ -0-	Antipsychotic
			Neuroleptic
			Sedative
(±) Tetrahydrocoptisine	-0-CH ₂ -0-	-0-CH ₂ -0-	DA ₂ agonist
			Monoamines depletor
			in brain
			Tranquilizer

Table 4 (continue)

(+) Tetrahydropalmatine *	OMe	OMe	OMe	OMe	CNS stimulant Dopamine depletor
					Inhibition of
					catecholamine uptake
(-) Tetrahydropalmatine *	ОМе	OMe	OMe	OMe	Analgesic
		1			Antiarrhythmic
					Catalepsic
			7.6		Dopaminergic
					antagonist
					Hyperthermic
			4		Hypnotic
					Increase the action
					of analgesic
					Increase the action
					of CNS deperssant
					Increase the thresho
					of hippocampal
					convulsive action
					Inhibition of
					catecholamine uptak
					Inhibition the
					activation of the
					reticular system

Table 4 (continue)

-) Tetrahydropalmatine*					Monoamine depletor
					in brain
					Sedative
					Spasmolytic
					Supperssed the
					cortical and
					subcortical reaction
					Supperssed the period
					of after affect
					Tranquillizer
(±) Tetrahydropalmatine	OMe	OMe	OMe	OMe	ACTH-like
					Antiarrhythmic
					Antimicrobial
					Ca ⁺² antagonist
					Decrease the speed of
					neuronal
					depolarization
					Hypotensive
					Monoamines depletor
					in brain
					Sedative
					Serotonergic antagonis
		1		. 1	

Table 4 (continue)

Tetrahydropalmatine	OMe	OMe	OMe	OMe	CNS depressant Inhibition of NADH oxidase Sedative Tranquillizer Uterotonic
(+) 2,3,9,10-Tetrahydroxy tetrahydroprotoberberine	ОН	ОН	ОН	ОН	Inhibition of dopamine- sensitive adenylate cyclase
(-) 2,3,9,10-Tetrahydroxy tetrahydroprotoberberine	ОН	ОН	ОН	ОН	Dopaminergic antagonist Inhibition of dopamine- sensitive adenylate cyclase
(±) 3,9,10-Trimethoxy tetrahydroprotoberbering	e	OMe	OMe	OMe	Tranquillizer

Tetrahydropseudoprotoberberines Structure Name 2 3 10 11 Activity (+) 10-Acetyl 2,3,11-OMe OMe OAc OMe | Analgesic trimethoxy Hypotensive tetrahydroprotoberberine Vasodilator Coreximine * OH OMe OMe OH Respirationy stimulant (Coramine) (-) 10,11-Dihydroxy OH Inhibition of dopamine OH tetrahydroprotoberberine sensitive adenylate . cyclase (\pm) 2,3-Dimethoxy 10,11--O-CH₂-O- Analgesic 0Me OMe methylenedioxy Hypotensive tetrahydroprotoberberine Vasodilator

Table 4 (continue)

+) 3-Hydroxy 10,11-dimethoxy tetrahydroprotoberberine	102	ОН	OMe	OMe	Hypotensive
+) 2-Methoxy 3,10,11- triĥydroxy tetrahydroprotoberberine	ОМе	ОН	ОН	ОН	Dopaminergic antagonist
+) 3-Methoxy 2,10,11- trihydroxy tetrahydroprotoberberine	ОН	OMe	ОН	ОН	Dopaminergic antagonist
(+) 11-Methoxy 2,3,10- trihydroxy tetrahydroprotoberberine	ОН	ОН	ОН	OMe	Dopaminergic antagonist
(±) 2,3-Methylenedioxy 9,10,11-trihydroxy tetrahydroprotoberberin		СН2-0	O- OH		H Inhibition of dopamine OH sensitive adenylate cyclase
(-) 2,3,10,11-Tetrahydroxy tetrahydroprotoberberin	OH	I 0	н О	Н	OH Dopaminergic antagonis Inhibition of monoamine oxidase

Table 4 (continue)

+) 2,3,10,11-Tetrahydroxy tetrahydroprotoberberine	ОН	ОН	ОН	ОН	Inhibition of - sensitive adenylate cyclase
,3,10,11-Tetrahydroxy tetrahydroprotoberberine	ОН	ОН	ОН	ОН	Inhibition of catecholamine uptake
±) 2,3,11-Trihydroxy tetrahydroprotoberberine	ОН	ОН	-	ОН	Inhibition of - sensitive adenylate cyclase Inhibition of dopamine- sensitive adenylate cyclase
(±) 3,10,11-Trihydroxy tetrahydroprotoberberine	-	ОН	ОН	ОН	Hypotensive
(-) Xylopinine*	OMe	OMe	OMe	OMe	Dopaminergic antagonis
(±) Xylopinine	OMe	OMe	OMe	OMe	Analgesic Hypotensive Vasodilator
Xylopinine	OMe	OMe	OMe	OMe	Antitussive Sympatholytic

Tetrahydroprotoberberines oxygenated at C-8

		Struc	ture				
Name	2	3	8	10	Activity		
±) 2,3,10-Trimethoxy 8-oxo tetrahydroprotoberberine	ОМе	OMe	0	OMe	Tranquillizer		

Tetrahydroprotoberberines methylated at carbon 13

			Stru				
Name	2	3	9	10	11	13	Activity
(+) Cavidine*	OMe	ОМе	-0-C	H ₂ -0-	-	Me	Spasmolytic
Corydaline*	OMe	OMe	OMe	ОМе	-	Ме	CNS depressant Uterotonic
2,3-Methylenedioxy 10,11-dimethoxy 13-methyl tetrahydro- protoberberine	-0-0	сн ₂ -о-	-	ОМе	OMe	Me	Cytotoxic

Table 4 (continue)

Ophiocarpine*	-0-CH ₂ -0-		OMe	OMe	-	Ме	Uterotonic	
(±) 13-Cianoxylopinine	О́Ме	OMe	-	OMe	OMe	CN	Hypotensive Vasodilator	
2,3-Methylenedioxy 10,11-dimethoxy 13-hydroxymethyl tetrahydro protoberberine	-0-0	H ₂ -0-	-	ОМе	OMe	СН2ОН	Cytotoxic	

Tertahydroprotoberberine with N-methylated

		\$	Struc				
Name	2	3	7	9	10	11	Activity
N-Methylisocorypalmine	ОН	ОМе	Ме	OMe	OMe	-	Positive inotropic
N-Methylxylopinine	ОМе	OMe	Me	-	ОМе	OMe	Curare-like

Name	1	2	3	4	9	10	11	12	Activeity
Capauridine*	ОН	ОМе	ОМе	-	ОМе	ОМе	-	-	Uterotonic
Capaurine*	ОН	0Me	0Me	-	OMe	ОН	-	-	Uterotonic
(+) 4-Formyl- xylopine	-	ОМе	ОМе	СНО		ОМе	OMe	-	Analgesic Hypotensive Sedative

Table 4 (continue)

(⁺)	2,3,10,11,12- pentamethoxy	-	OMe	OMe	-	-	OMe	OMe	OMe	Tranquillizer
	tetrahydro- protoberberine						•			
(+)	1,2,3,10- tetramethoxy	OMe	OMe	OMe	-	-	OMe	-	-	Tranquillizer
	tetrahydro- protoberberine									
(+)	1,3,10- trimethoxy	OMe	-	OMe	-	-	ОМе	-	-	Tranquillize
	tetrahydro- protoberberine									
(±)	3,4,10- trimethoxy	-	-	OMe	OMe	-	OMe	-	-	Tranquillize
	tetrahydro- protoberberine									

4.2 Pharmacology of aprophine alkaloids

The aporphine alkaloids display a wide range of pharmacologic activities, although none are commercial items.

1,2 - Methylenedioxyaporphine increases arterial blood pressure, but higher doses cause strychnine - like convulsions. The methohydroxide salt has a curare - like action. (Cordell, 1981)

Isothebaine increased intestinal muscle tone in rabbits and also amplified uterine contractions in the rat. Other activities observed include decreased motor activity and analgesia (mice), and an anti-inflammatory effect(rats). (Cordell, 1981) and depress central nervous system. (Shamma, 1972)

Glaucine and dicentrine cause narcosis in animals, and with larger doses convulsions. (Shamma, 1972). Glaucine reduced blood pressure and inhibited respiration in cats and had antitussive effects resembling codeine, but of longer duration.

In rats and cats a potentially useful hypoglycemic effect was observed at 12-mg/kg doses. Dehydroglaucine has antibacterial activity. (Cordell, 1981)

Corydine has central nervous system depressant and hypotensive activity and blocks transmission of nerve impulse. The corresponding 11-demethyl derivative, corytuberine, accelerates respiration and stimulates secretions.

Bulbocapnine antagonizes the effects of apomorphine and amphetamine, depresses the central nervous system, and causes catalepsy in mice. Xylopine has sedative and analgesic activity and isoboldine is an insect-feeding inhibitor.

Apomorphine although not a natural product has been quite well studied, it has hypotensive activity and is a powerful emetic, suitable for rapid emesis after ingestion of poisons. Of more interest from a therapeutic point of view is its stimulation of the dopaminergic system in rats and mice, and consequently its potential anti-Parkinsonism activity. Also of interest are reports that it can decrease serum prolactin levels. (Cordell, 1981)

Apocodeine may have useful emetic activity. Boldine is only slightly toxic and does not cause addiction. It has mild sedative, diuretic, and antispasmodic action, and also increases the secretions of the liver and salivary glands. Laurifoline chloride has some hypotensive activity, while corytuberine accelerates respiration and slow the pulse. Xylopine is supposed to possess sedative and analgesic activity. (Shamma, 1972)